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- (54) Amino-hydroxy-methyl-isoxazole-propionate binding human glutamate receptors.
- ⑤ Described herein are isolated polynucleotides which code for a family of AMPA-type human CNS receptors. The receptors are characterized structurally and the construction and use of cell lines expressing these receptors are disclosed.

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F/G /6  CEMPICANTAGETTICAAAATICTCATCTCTCTCTCOAAACTICTCTCT  0	FIG. W  CENTRAL CONTROL MAN CONTROL MAN CONTROL MAN CONTROL CO

#### Background of the Invention

#### Field of the Invention

This invention is concerned with applications of recombinant DNA technology in the field of neurobiology. More particularly, the invention relates to the cloning and expression of DNA coding for excitatory amino acid (EAA) receptors, especially human EAA receptors.

#### Background of the Invention

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In the mammalian central nervous system (CNS), the transmission of nerve impulses is controlled by the interaction between a neurotransmitter substance released by the "sending" neuron which then binds to a surface receptor on the "receiving" neuron to cause excitation thereof. L-glutamate is the most abundant neurotransmitter in the CNS, and mediates the major excitatory pathway in vertebrates. Glutamate is therefore referred to as an excitatory amino acid (EAA) and the receptors which respond to it are variously referred to as glutamate receptors, or more commonly as EAA receptors.

Using tissues isolated from mammalian brain, and various synthetic EAA receptor agonists, knowledge of EAA receptor pharmacology has been refined somewhat. Members of the EAA receptor family are now grouped into three main types based on differential binding to such agonists. One type of EAA receptor, which in addition to glutamate also binds the agonist NMDA (N-methyl-D-aspartate), is referred to as the NMDA type of EAA receptor. Two other glutamate-binding types of EAA receptor, which do not bind NMDA, are named according to their preference for binding with two other EAA receptor agonists, namely AMPA ( $\alpha$ -amino-3-hydroxy-5-methyl-isoxazole-4-propionate), and kainate. Particularly, receptors which bind glutamate but not NMDA, and which bind with greater affinity to kainate than to AMPA, are referred to as kainate type EAA receptors. Similarly, those EAA receptors which bind glutamate but not NMDA, and which bind AMPA with greater affinity than kainate are referred to as AMPA type EAA receptors.

The glutamate-binding EAA receptor family is of great physiological and medical importance. Glutamate is involved in many aspects of long-term potentiation (learning and memory), in the development of synaptic plasticity, in epileptic seizures, in neuronal damage caused by ischemia following stroke or other hypoxic events, as well as in other forms of neurodegenerative processes. However, the development of therapeutics which modulate these processes has been very difficult, due to the lack of any homogeneous source of receptor material with which to discover selectively binding drug molecules, which interact specifically at the interface of the EAA receptor. The brain derived tissues currently used to screen candidate drugs are heterogeneous receptor sources, possessing on their surface many receptor types which interfere with studies of the EAA receptor/ligand interface of interest. The search for human therapeutics is further complicated by the limited availability of brain tissue of human origin. It would therefore be desirable to obtain cells that are genetically engineered to produce only the receptor of interest. With cell lines expressing cloned receptor genes, a substrate which is homogeneous for the desired receptor is provided, for drug screening programs.

Recently, genes encoding substituent polypeptides of EAA receptors from non-human sources, principally rat, have been discovered. Hollmann et al., Nature 342: 643, 1989 described the isolation from rat of a gene referred to originally as GluR-K1 (but now called simply GluR1). This gene encodes a member of the rat EAA receptor family, and was originally suspected as being of the kainate type. Subsequent studies by Keinanen et al., Science 249: 556, 1990, showed, again in rat, that a gene called GluR-A, which was identical to the previously isolated GluR1, in fact encodes a receptor not of the kainate type, but rather of the AMPA type. These two groups of researchers have since reported as many as five related genes isolated from rat sources. Boulter et al., Science 249: 1033, 1990, revealed that, in addition to GluR1, the rat contains 3 other related genes, which they called GluR2, GluR3, and GluR4, and Bettler et al., Neuron 5: 583. 1990 described GluR5. Keinanen et al., supra, described genes called GluR-A, GluR-B, GluR-C and GluR-D which correspond precisely to GluR1, GluR2, GluR3 and GluR4 respectively. Sommer et al., Science 249: 1580, 1990 also showed, for GluR-A, GluR-B, GluR-C and GluR-D two alternatively spliced forms for each gene. These authors, as well as Monyer et al., Neuron 6: 799, 1991 were able to show that the differently spliced versions of these genes are differentially expressed in the rat brain.

There has emerged from these molecular cloning advances a better understanding of the structural features of EAA receptors and their subunits, as they exist in the rat brain. According to the current model of EAA receptor structure, each is heteromenic in structure, consisting of individual membrane-anchored subunits, each having four transmembrane regions, and extracellular domains that dictate ligand binding properties to some extent and contribute to the ion-gating function served by the receptor complex. Keinanen et al, supra, have shown for example that each subunit of the rat GluR receptor, including those designated GluR-A, GluR-

B, GluR-C and GluR-D, display cation channel activity gated by glutamate, by AMPA and by kainate, in their unitary state. When expressed in combination however, for example GluR-A in combination with GluR-B, gated ion channels with notably larger currents are produced by the host mammalian cells.

In the search for therapeutics useful to treat CNS disorders in humans, it is highly desirable of course to provide a screen for candidate compounds that is more representative of the human situation than is possible with the rat receptors isolated to date. It is particularly desirable to provide cloned genes coding for human receptors, and cell lines expressing those genes, in order to generate a proper screen for human therapeutic compounds. These, accordingly, are objects of the present invention.

#### Summary of the Invention

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The present invention provides isolated polynucleotides that code for a family of AMPA-binding human EAA receptors, herein referred to as "GluR receptors". By providing polynucleotides that code specifically for CNS receptors native to humans, the present invention provides means for evaluating the human nervous system, and particularly for assessing potentially therapeutic interactions between the AMPA-binding human EAA receptors and selected natural and synthetic ligands.

In one of its aspects, the present invention provides an isolated polynucleotide that codes for an EAA receptor belonging to the human GluR family. Alternatively, the polynucleotide may code for an AMPA-binding fragment of a human GluR receptor, or for an AMPA-binding variant of a human GluR receptor. According to specific embodiments of the present invention, the isolated polynucleotide encodes the human GluR1B receptor, the amino acid sequence of which is identified in Figure 1 (SEQ ID NO: 2), the human GluR2B receptor the amino acid sequence of which is identified in Figure 2 (SEQ ID NO: 4), and the human GluR3A receptor, the amino acid sequence of which is identified in Figure 3 (SEQ ID NO: 6). According to another embodiment of the invention, the polynucleotide encodes an AMPA-binding variant of the human GluR receptor. One such variant is identified herein as the human GluR3B receptor, the amino acid sequence of which is identified in Figure 4 (SEQ ID NO: 8). In various specific embodiments of the present invention, the polynucleotide consists of DNA e.g. cDNA, or of RNA e.g. messenger RNA. In other embodiments of the present invention, the polynucleotide may be coupled to a reporter molecule, such as a radioactive label, for use in autoradiographic studies of human GluR receptor tissue distribution. In further embodiments of the present invention, fragments of the polynucleotides of the invention, including radiolabelled versions thereof, may be employed either as probes for detection of glutamate receptor-encoding polynucleotides, as primers appropriate for amplifying such polynucleotides present in a biological specimen, or as templates for expression of a GluR receptor or AMPA-binding fragments or variants thereof.

According to another aspect of the present invention, there is provided a cellular host that produces an AMPA-type human glutamate receptor, and is characterized by the incorporation therein of a polynucleotide of the present invention. In embodiments of the present invention, the polynucleotide is a DNA molecule and is incorporated for expression and secretion in the cellular host, to yield, upon culturing, a functional, membrane-bound human GluR receptor. In other embodiments of the present invention, the polynucleotide is an RNA molecule which is introduced into the cellular host to yield a human GluR receptor as a functional, membrane-bound product of translation.

According to another aspect of the invention, there is provided a process for obtaining a substantially homogeneous source of a human EAA receptor useful for performing ligand binding assays, which comprises the steps of culturing a genetically engineered cellular host of the invention, and then recovering the cultured cells. Optionally, the cultured cells may be treated to obtain membrane preparations thereof, for use in the ligand binding assays.

According to another aspect of the present invention, there is provided a method for assaying interaction between a test ligand and a human EAA receptor, comprising the steps of incubating the test ligand under appropriate conditions with a human GluR receptor source, i.e., a cellular host of the invention or a membrane-preparation derived therefrom, and then determining the extent or result of binding between the substance and the receptor source.

These and other aspects of the invention are now described in greater detail with reference to the accompanying drawings, in which:

#### Brief Description of the Drawings

Figure 1 provides a DNA sequence coding for the human GluR1B receptor, and the amino acid sequence thereof (SEQ ID NOS: 1 and 2);

Figure 2 provides a DNA sequence coding for the human GluR2B receptor, and the amino acid sequence

thereof (SEQ ID NOS: 3 and 4);

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Figure 3 provides a DNA sequence coding for the human GluR3A receptor, and the amino acid sequence thereof (SEQ ID NOS: 5 and 6);

Figure 4 provides a DNA sequence coding for the human GluR3B receptor, and the amino acid sequence thereof (SEQ ID NOS: 7 and 8);

Figure 5 provides the amino acid sequence of the human GluR3A receptor (SEQ ID NO: 9) and the human GluR3B receptor (SEQ ID NO: 10) in a region of dissimilarity;

Figure 6 depicts the strategy employed in cloning the human GluR3A receptor-encoding DNA illustrated in Figure 3;

Figure 7 depicts the strategy employed in cloning the human GluR3B receptor-encoding DNA illustrated in Figure 4;

Figure 8 depicts the strategy employed in generating recombinant DNA expression constructs incorporating the GluR3A receptor-encoding DNA;

15 Figure 9 depicts the strategy employed in generating recombinant DNA expression constructs incorporating the GluR1B receptor-encoding DNA (SEQ ID NOS: 11 and 12 are also shown in this figure);

Figure 10 depicts the strategy employed in cloning the human GluR2B receptor-encoding DNA illustrated in Figure 2;

Figure 11 depicts the strategy employed in generating recombinant DNA expression constructs incorporating the GluR2B receptor-encoding DNA;

Figure 12 illustrates the AMPA-binding property of the human GluR1B receptor,

Figure 13 illustrates the AMPA-binding property of the human GluR2B receptor,

Figure 14 illustrates the AMPA-binding property of the human GluR3A receptor;

Figures 15 & 16 illustrate a Scatchard analysis of human GluR1B and GluR2B receptor AMPA binding; and Figure 17 graphically shows AMPA competition binding data for the GluR2B receptor.

#### **Detailed Description of the Preferred Embodiments**

The invention relates to human CNS receptors of the AMPA-binding type, and is directed more particularly to novel receptors belonging to a family herein referred to as "GluR receptors", and provides isolated polynu-ficeotides that code for such receptors. The term "isolated" is used herein with reference to intact polynucleotides that are generally less than about 4,000 nucleotides in length and which are otherwise isolated from DNA coding for other human proteins.

As used herein, the term "GluR receptors" is intended to embrace the human GluR1B, GluR2B and GluR3A receptors, AMPA-binding variants related thereto, as well as AMPA-binding fragments of the GluR1B, GluR2B and GluR3A receptors. Receptor variants within the scope of the present invention are functional variants of a parent receptor, i.e., one of GluR1B, GluR2B, GluR3A and GluR3B, which include conservative amino acid substitutions.

The term "AMPA-binding", as used herein with respect to receptors, and variants and fragments thereof, refers to a ligand binding profile which reveals glutamate binding and relative greater binding affinity for AMPA than for either glutamate, kainate or NMDA, as determined using assays of conventional design, such as the assays herein described.

In the present specification, an AMPA-binding receptor is said to be "functional" if a cellular host producing it exhibits *de novo* channel activity when exposed appropriately to AMPA, as determined by the established electrophysiological assays described for example by Hollmann et al., *supra*, or by any other assay appropriate for detecting conductance across a cell membrane.

Members of the human GluR family of the invention possess structural features characteristic of the EAA receptors in general, including extracellular N- and C-terminal regions, as well as four internal hydrophobic domains which serve to anchor the receptor within the cell surface membrane.

More specifically, the GluR1B receptor is a protein characterized structurally as a single polypeptide chain that is produced initially in precursor form bearing an 18 amino acid residue N-terminal signal peptide, and is transported to the cell surface in mature form, lacking the signal peptide and consisting of 888 amino acids arranged in the sequence illustrated, by single letter code, in Figure 1 (SEQ ID NOS: 1 and 2). Unless otherwise stated, the term human GluR receptor, either generally or with reference to a particular member of the receptor family, refers to the mature form of the receptor. Thus, the amino acid residues of these receptors are numbered in Figures 1-4 with reference to the mature protein sequence. With respect to structural domains of the GluR1B receptor, hydropathy analysis reveals four putative transmembrane domains, one spanning residues 521-540 inclusive (TM-1), another spanning residues 567-585 (TM-2), a third spanning residues 596-614 (TM-3) and the fourth spanning residues 788-808 (TM-4). Based on this assignment, it is likely that the human

GluR1B receptor structure, in its natural membrane-bound form, consists of a 520 amino acid N-terminal extracellular domain, followed by a hydrophobic region containing four transmembrane domains and an extracellular, 80 amino acid C-terminal domain.

The GluR2B receptor, in precursor form bears a 21 amino acid residue N-terminal signal peptide, and in mature form, consists of 862 amino acids arranged in the sequence illustrated, by single letter code, in Figure 2 (SEQ ID NOS: 3 and 4). With respect to structural domains of the receptor, hydropathy analysis reveals four putative transmembrane domains, one spanning residues 525-544 inclusive (TM-1), another spanning residues 571-589 (TM-2), a third spanning residues 600-618 (TM-3) and the fourth spanning residues 792-812 (TM-4). Based on this assignment, it is likely that the human GluR2B receptor structure, in its natural membrane-bound form, consists of a 524 amino acid N-terminal extracellular domain, followed by a hydrophobic region containing four transmembrane domains and an extracellular, 50 amino acid C-terminal domain.

The GluR3A member of the human GluR family bears a 22 amino acid residue N-terminal signal peptide in precursor form, and is transported to the cell surface in mature form, lacking the signal peptide and consisting of 866 amino acids arranged in the sequence illustrated, by single letter code, in Figure 3 (SEQ ID NOS: 5 and 6). The four putative transmembrane domains of the GluR3A receptor are as follows: one spans residues 527-546 inclusive (TM-1), another spans residues 575-593 (TM-2), a third spans residues 604-622 (TM-3) and the fourth spans residues 796-816 (TM-4). Based on this assignment, it is likely that the human GluR3A receptor structure, in its natural membrane-bound form, consists of a 526 amino acid N-terminal extracellular domain, followed by a hydrophobic region containing four transmembrane domains and an extracellular, 50 amino acid C-terminal domain.

Structurally related variants of the GluR parent receptors identified above also exist. Specifically, a structurally related variant of the human GluR3A receptor, namely the GluR3B receptor, has also been identified. This variant occurs naturally in human brain tissue, and like GluR3A, the GluR3B receptor is 866 amino acids in length, as shown in Figure 4 (SEQ ID NOS: 7 and 8), in its mature, membrane-bound form. The GluR3B receptor initially bears a signal peptide identical to that borne on the GluR3A receptor. Four transmembrane domains are also apparent from the GluR3B sequence, and indicate that these domains lie in the same amino acid regions identified in connection with the GluR3A receptor.

With respect to primary structure, the human GluR3B receptor differs from the GluR3A receptor in a 36 amino acid region separating transmembrane domains TM-3 and TM-4, i.e. residues 748-783. For comparison, the sequences of GluR3A and GluR3B in this region are compared in Figure 5 (SEQ ID NOS: 9 and 10).

Binding assays performed with various ligands, and with membrane preparations derived from mammalian cells engineered genetically to produce the human GluR receptors in membrane-bound form indicate that the human GluR receptors bind selectively to AMPA, relative particularly to kainate and NMDA. This feature, coupled with the medically significant connection between AMPA-type receptors and neurological disorders and disease indicate that the present receptors, as well as AMPA-binding fragments and variants thereof, will serve as valuable tools in the screening and discovery of ligands useful to modulate *in vivo* interactions between such receptors and their natural ligand, glutamate. Thus, a key aspect of the present invention resides in the construction of cells that are engineered genetically to produce a human GluR receptor, to serve as a ready and homogeneous source of receptor for use in *vitro* ligand binding and/or channel activation assays.

For use in the ligand binding assays, it is desirable to construct by application of genetic engineering techniques a host cell, either prokaryotic or eukaryotic, that produces a human GluR receptor as a heterologous and membrane-bound product. According to one embodiment of the invention, the construction of such engineered cells is achieved by introducing into a selected host cell a recombinant DNA construct in which DNA coding for a secretable form of the desired human GluR receptor, i.e., a form bearing its native signal peptide or a functional, heterologous equivalent thereof, is linked operably with expression controlling elements that are functional in the selected host to drive expression of the receptor-encoding DNA, and thus elaborate the desired human GluR receptor protein. Such cells are herein characterized as having the receptor-encoding DNA incorporated "expressibly" therein. The receptor-encoding DNA is referred to as "heterologous" with respect to the particular cellular host if such DNA is not naturally found in the particular host. The particular cell type selected to serve as host for production of the human GluR receptor can be any of several cell types currently available in the art, including both prokaryotic and eukaryotic cells, but should not of course be a cell type that in its natural state elaborates a surface receptor that can bind excitatory amino acids, and so confuse the assay results sought from the engineered cell line. Generally, such problems are avoided by selecting as host a non-neuronal cell type, and can further be avoided using non-human cell lines, as is conventional. It will be appreciated that neuronal- and human-type cells may nevertheless serve as expression hosts, provided that "background" binding to the test ligand is accounted for in the assay results.

According to one embodiment of the present invention, the cell line selected to serve as host for human GluR receptor production is a mammalian cell. Several types of such cell lines are currently available for ge-

netic-engineering work, and these include the chinese hamster ovary (CHO) cells for example of K1 lineage (ATCC CCL 61) including the Pro5 variant (ATCC CRL 1281); the fibroblast-like cells derived from SV40-transformed African Green monkey kidney of the CV-1 lineage (ATCC CCL 70), of the COS-1 lineage (ATCC CRL 1650) and of the COS-7 lineage (ATCC CRL 1651); murine L-cells, murine 3T3 cells (ATCC CRL 1658), murine C127 cells, human embryonic kidney cells of the 293 lineage (ATCC CRL 1573), human carcinoma cells including those of the HeLa lineage (ATCC CCL 2), and neuroblastoma cells of the lines IMR-32 (ATCC CCL 127), SK-N-MC (ATCC HTB 10) and SK-N-SH (ATCC HTB 11).

A variety of gene expression systems have been adapted for use with these hosts and are now commercially available, and any one of these systems can be selected to drive expression of human GluR receptorencoding DNA. These systems, available typically in the form of plasmidic vectors, incorporate expression cassettes the functional components of which include DNA constituting expression controlling sequences, which are host-recognized and enable expression of the receptor-encoding DNA when linked 5' thereof. The systems further incorporate DNA sequences which terminate expression when linked 3' of the receptor-encoding region. Thus, for expression in the selected mammalian cell host, there is generated a recombinant DNA expression construct in which DNA coding for a secretable form of the receptor is linked with expression controlling DNA sequences recognized by the host, and which include a region 5' of the receptor-encoding DNA to drive expression, and a 3' region to terminate expression. The plasmidic vector harboring the recombinant DNA expression construct typically incorporates such other functional components as an origin of replication, usually virally-derived, to permit replication of the plasmid in the expression host and desirably also for plasmid amplification in a bacterial host, such as E.coli. To provide a marker-enabling selection of stably transformed recombinant cells, the vector will also incorporate a gene conferring some survival advantage on the transformants, such as a gene coding for neomycin resistance in which case the transformants are plated in medium supplemented with neomycin.

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Included among the various recombinant DNA expression systems that can be used to achieve mammalian cell expression of the receptor-encoding DNA are those that exploit promoters of viruses that infect mammalian cells, such as the promoter from the cytomegalovirus (CMV), the Rous sarcoma virus (RSV), simian virus (SV40), murine mammary tumor virus (MMTV) and others. Also useful to drive expression are promoters such as the LTR of retroviruses, insect cell promoters such as those regulated by temperature, and isolated from Drosophila, as well as mammalian gene promoters such as those regulated by heavy metals i.e.the metalothionein gene promoter, and other steroid-inducible promoters.

For incorporation into the recombinant DNA expression vector, DNA coding for a selected human GluR receptor, e.g. one of the human GluR1B, GluR2B or GluR3A receptors, or an AMPA-binding fragment or variant thereof, e.g. GluR3B, can be obtained by applying selected techniques of gene isolation or gene synthesis. As described in more detail in the examples herein, human GluR receptors are encoded within the genome of human brain tissue, and can therefore be obtained from human DNA libraries by careful application of conventional gene isolation and cloning techniques. This typically will entail extraction of total messenger RNA from a fresh source of human brain tissue, preferably cerebellum or hippocampus tissue, followed by conversion of message to cDNA and formation of a library in for example a bacterial plasmid, more typically a bacteriophage. Such bacteriophage harboring fragments of the human DNA are typically grown by plating on a lawn of susceptible E. coli bacteria, such that individual phage plaques or colonies can be isolated. The DNA carried by the phage colony is then typically immobilized on a nitrocellulose or nylon-based hybridization membrane, and then hybridized, under carefully controlled conditions, to a radioactively (or otherwise) labelled oligonucleotide probe of appropriate sequence to identify the particular phage colony carrying receptor-encoding DNA or fragment thereof. It will be understood, for example, that selective hybridization, i.e. hybridization of a DNA sequence that is completely complementary to the probe, will be conducted under stringent hybridization conditions. Typically, the gene or a portion thereof so identified is subcloned into a plasmidic vector for nucleic acid sequence analysis.

In specific embodiments of the invention, the GluR1B receptor is encoded by the DNA sequence illustrated in Figure 1 (SEQ ID NO: 1), the GluR2B receptor is encoded by the DNA sequence illustrated in Figure 2 (SEQ ID NO: 3) and the GluR3A and GluR3B receptors are encoded by the DNA sequences illustrated respectively in Figures 3 (SEQ ID NO: 5) and 4 (SEQ ID NO: 7). Alternatively, codons within the illustrated DNA sequences coding for the GluR receptors may be replaced by synonymous codon equivalents, such synonymous codon replacements being well-known in the art.

The illustrated DNA sequences constitute cDNA sequences identified in human brain cDNA libraries in the manner exemplified herein. Having herein provided the nucleotide sequence of various members of the human GluR receptor family, however, it will be appreciated that polynucleotides encoding the receptors can be obtained by other routes. Automated techniques of gene synthesis and/or amplification can be performed to generate DNA coding therefor. Because of the length of the human GluR receptor-encoding DNA, application of

automated synthesis may require staged gene construction, in which regions of the gene up to about 300 nucleotides in length are synthesized individually and then ligated in correct succession by overhang complementarity for final assembly. Individually synthesized gene regions can be-amplified prior to assembly, using established polymerase chain reaction (PCR) technology.

By the application of automated gene synthesis techniques, there is provided a means to generate polynucleotides that encode variants of naturally occurring human GluR receptors, i.e. GluR1B, GluR2B, GluR3A and GluR3B. It will be appreciated, for example, that polynucleotides coding for the human GluR receptors herein described can be generated by substituting synonymous codons for those represented in the naturally occurring polynucleotide sequences herein identified. In addition, polynucleotides coding for human GluR receptor variants can be generated which for example incorporate one or more e.g. 1-10, single amino acid substitutions, deletions or additions. Since it will for the most part be desirable to retain the natural ligand binding profile of the receptor for screening purposes, it is desirable to limit amino acid substitutions, for example to the so-called conservative replacements in which amino acids of like charge are substituted, and to limit substitutions to those sites less critical for receptor activity e.g. within about the first 20 N-terminal residues of the mature receptor, and such other regions as are elucidated upon receptor domain mapping.

With appropriate template DNA in hand, the technique of PCR amplification may also be used to directly generate all or part of the final gene. In this case, primers are synthesized which will prime the PCR amplification of the final product, either in one piece, or in several pieces that may be ligated together. This may be via step-wise ligation of blunt ended, amplified DNA fragments, or preferentially via step-wise ligation of fragments containing naturally occurring restriction endonuclease sites. In this application, it is possible to use either cDNA or genomic DNA as the template for the PCR amplification. In the former case, the cDNA template can be obtained from commercially available or self-constructed cDNA libraries of various human brain tissues, including hippocampus and cerebellum.

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Once obtained, the receptor-encoding DNA is incorporated for expression into any suitable expression vector, and host cells are transfected therewith using conventional procedures, such as DNA-mediated transformation, electroporation, or particle gun transformation. Expression vectors may be selected to provide transformed cell lines that express the receptor-encoding DNA either transiently or in a stable manner. For transient expression, host cells are typically transformed with an expression vector harboring an origin of replication functional in a mammalian cell. For stable expression, such replication origins are unnecessary, but the vectors will typically harbour a gene coding for a product that confers on the transformants a survival advantage, to enable their selection. Genes coding for such selectable markers include the E. coli *gpt* gene which confers resistance to mycophenolic acid, the *neo* gene from transposon Tn5 which confers resistance to the antibiotic G418 and to neomycin, the *dhfr* sequence from murine cells or E. coli which changes the phenotype of DHFR-cells into DHFR+ cells, and the *tk* gene of herpes simplex virus, which makes TK- cells phenotypically TK+ cells. Both transient expression and stable expression can provide transformed cell lines, and membrane preparations derived therefrom, for use in ligand screening assays.

For use in screening assays, cells transiently expressing the receptor-encoding DNA can be stored frozen for later use, but because the rapid rate of plasmid replication will lead ultimately to cell death, usually in a few days, the transformed cells should be used as soon as possible. Such assays may be performed either with intact cells, or with membrane preparations derived from such cells. The membrane preparations typically provide a more convenient substrate for the ligand binding experiments, and are therefore preferred as binding substrates. To prepare membrane preparations for screening purposes, i.e., ligand binding experiments, frozen intact cells are homogenized while in cold water suspension and a membrane pellet is collected after centrifugation. The pellet is then washed in cold water, and dialyzed to remove endogenous EAA ligands such as glutamate, that would otherwise compete for binding in the assays. The dialyzed membranes may then be used as such, or after storage in lyophilized form, in the ligand binding assays. Alternatively, intact, fresh cells harvested about two days after transient transfection or after about the same period following fresh plating of stably transfected cells, can be used for ligand binding assays by the same methods as used for membrane preparations. When cells are used, the cells must be harvested by more gentle centrifugation so as not to damage them, and all washing must be done in a buffered medium, for example in phosphate-buffered saline, to avoid osmotic shock and rupture of the cells.

The binding of a substance, i.e., a candidate ligand, to a human GluR receptor of the invention is evaluated typically using a predetermined amount of cell-derived membrane (measured for example by protein determination), generally from about 25µg to 100µg. Generally, competitive binding assays will be useful to evaluate the affinity of a test compound relative to AMPA. This competitive binding assay can be performed by incubating the membrane preparation with radiolabelled AMPA, for example [3H]-AMPA, in the presence of unlabelled test compound added at varying concentrations. Following incubation, either displaced or bound radiolabelled AMPA can be recovered and measured, to determine the relative binding affinities of the test compound

and AMPA for the particular receptor used as substrate. In this way, the affinities of various compounds for the AMPA-binding human EAA receptors can be measured. Alternatively, a radiolabelled analogue of glutamate may be employed in place of radiolabelled AMPA, as competing ligand.

As an alternative to using cells that express receptor-encoding DNA, ligand characterization may also be performed using cells for example Xenopus oocytes, that yield functional membrane-bound receptor following introduction by injection either of receptor-encoding messenger RNA into the oocyte cytoplasm, or of receptor-encoding DNA into the oocyte nucleus. To generate the messenger RNA of cytoplasmic delivery, the receptor-encoding DNA is typically subcloned first into a plasmidic vector adjacent a suitable promoter region, such as the T3 or T7 bacteriophage promoters, to enable transcription into RNA message. RNA is then transcribed from the inserted gene *in vitro*, collected and then injected into Xenopus oocytes. Following the injection of nL volumes of an RNA solution, the oocytes are left to incubate for up to several days, and are then tested for the ability to respond to a particular ligand molecule supplied in a bathing solution. Since functional EAA receptors act in part by operating a membrane channel through which ions may selectively pass, the functioning of the receptor in response to a particular ligand molecule in the bathing solution may typically be measured as an electrical current utilizing microelectrodes inserted into the cell, in the established manner.

In addition to using the receptor-encoding DNA to construct cell lines useful for ligand screening, expression of the DNA can, according to another aspect of the invention, be performed to produce AMPA-binding fragments of the receptor in soluble form, for structure investigation, to raise antibodies and for other experimental uses. It is expected that the portion of the human GluR receptor responsible for AMPA-binding resides on the outside of the cell, i.e., is extracellular. It is therefore desirable in the first instance to facilitate the characterization of the receptor-ligand interaction by providing this extracellular ligand-binding domain in quantity and in isolated form, i.e., free from the remainder of the receptor. To accomplish this, the full-length human GluR receptor-encoding DNA may be modified by site-directed mutagenesis, so as to introduce a translational stop codon into the extracellular N-terminal region, immediately before the sequence encoding the first transmembrane domain (TM1), i.e., before residue 521 of GluR1B, before residue 525 in GluR2B, or before residue 527 of GluR3A and GluR3B. Since there will no longer be produced any transmembrane domain(s) to "anchor" the receptor into the membrane, expression of the modified gene will result in the secretion, in soluble form, of only the extracellular ligand-binding domain. Standard ligand-binding assays may then be performed to ascertain the degree of binding of a candidate compound to the extracellular domain so produced. It may of course be necessary, using site-directed mutagenesis, to produce several different versions of the extracellular regions, in order to optimize the degree of ligand binding to the isolated domains.

For use in ligand binding assays according to the present invention, AMPA-binding fragments of the receptor will first be anchored to a solid support using any one of various techniques. In one method, the C-terminal end of the receptor peptide fragment may be coupled to a derivatized, insoluble polymeric support, for example, cross-linked polystyrene or polyamide resin. Once anchored to the solid support, the frament is useful to screen candidate ligands for receptor binding affinity. For this purpose, competition-type ligand-binding assays, as described above using full-length receptor, are commonly used. Fragments secured to a solid support are bound with a natural ligand, i.e. AMPA, in the presence of a candidate ligand. One of AMPA or candidate ligand is labelled, for example radioactively, and following a suitable incubation period, the degree of AMPA displacement is determined by measuring the amount of bound or unbound label.

Alternatively, it may be desirable to produce an extracellular domain of the receptor which is not derived from the amino-terminus of the mature protein, but rather from the carboxy-terminus instead, for example domains immediately following the fourth transmembrane domain (TM4), i.e., residing between amino acid residues 809-888 of GluR1B, residues 813-862 of GluR2B, or residues 817-866 of GluR3A or GluR3B. In this case, site-directed mutagenesis and/or PCR-based amplification techniques may readily be used to provide a defined fragment of the gene encoding the receptor domain of interest. Such a DNA sequence may be used to direct the expression of the desired receptor fragment, either intracellularly, or in secreted fashion, provided that the DNA encoding the gene fragment is inserted adjacent to a translation start codon provided by the expression vector, and that the required translation reading frame is carefully conserved.

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It will be appreciated that the production of such AMPA-binding fragments of a GluR receptor may be accomplished in a variety of host cells. Mammalian cells such as CHO cells may be used for this purpose, the expression typically being driven by an expression promoter capable of high-level expression, for example the CMV (cytomegalovirus) promoter. Alternately, non-mammalian cells, such as insect Sf9 (Spodoptera frugiperda) cells may be used, with the expression typically being driven by expression promoters of the baculovirus, for example the strong, late polyhedrin protein promoter. Filamentous fungal expression systems may also be used to secrete large quantities of such extracellular domains of the EAA receptor. Aspergillus nidulans, for example, with the expression being driven by the alcA promoter, would constitute such an acceptable system. In addition to such expression hosts, it will be further appreciated that any prokaryotic or other eukaryotic ex-

pression system capable of expressing heterologous genes or gene fragments, whether intracellularly or extracellularly would be similarly acceptable.

For use particularly in detecting the presence and/or location of a human GluR receptor, for example in brain tissue, the present invention also provides, in another of its aspects, labelled antibody to a human GluR receptor. To raise such antibodies, there may be used as immunogen either the intact, soluble receptor or an immunogenic fragment thereof i.e. a fragment capable of eliciting an immune response, produced in a microbial or mammalian cell host as described above or by standard peptide synthesis techniques. Regions of human GluR receptor particularly suitable for use as immunogenic fragments include those corresponding in sequence to an extracellular region of the receptor, or a portion of the extracellular region. For example, peptides consisting of residues 1-526 of the GluR3A receptor or a fragment thereof comprising at least about 10 residues, including particularly fragments containing residues 178-193 or 479-522; and peptides corresponding to the region between transmembrane domains TM-2 and TM-3 of the GluR3A receptor, such as a peptide consisting of residues 594-603. Peptides consisting of the C-terminal domain (residues 817-866 of the GluR3A receptor), or fragment thereof, may also be used for the raising of antibodies.

The raising of antibodies to the selected human GluR receptor or immunogenic fragment can be achieved, for polyclonal antibody production, using immunization protocols of conventional design, and any of a variety of mammalian hosts, such as sheep, goats and rabbits. Alternatively, for monoclonal antibody production, immunocytes such as splenocytes can be recovered from the immunized animal and fused, using hybridoma technology, to a myeloma cells. The fusion products are then screened by culturing in a selection medium, and cells producing antibody are recovered for continuous growth, and antibody recovery. Recovered antibody can then be coupled covalently to a detectable label, such as a radiolabel, enzyme label, luminescent label or the like, using linker technology established for this purpose.

In detectably labelled form, e.g. radiolabelled form, DNA or RNA coding for a human GluR receptor, and selected regions thereof, may also be used, in accordance with another aspect of the present invention, as hybridization probes for example to identify sequence-related genes resident in the human or other mammalian genomes (or cDNA libraries) or to locate the human GluR-encoding DNA in a specimen, such as brain tissue. This can be done using either the intact coding region, or a fragment thereof having radiolabelled e.g. <sup>32</sup>P, nucleotides incorporated therein. To identify the human GluR-encoding DNA in a specimen, it is desirable to use either the full length cDNA coding therefor, or a fragment which is unique thereto. With reference to Figures 1-4 (SEQ ID NOS: 1-8), such nucleotide fragments include those comprising at least about 17 nucleic acids, and otherwise corresponding in sequence to a region coding for an extracellular N-terminal or C-terminal region of the receptor, or representing a 5'-untranslated or 3'-untranslated region thereof. Such oligonucleotide sequences, and the intact gene itself, may also be used of course to clone human GluR-related human genes, particularly cDNA equivalents thereof, by standard hybridization techniques.

Embodiments of the present invention are described in detail in the following specific examples which are not to be construed as limiting:

#### Example 1 - Isolation of DNA coding for the human GluR3A receptor

The particular strategy used to clone the human GluR3A receptor is depicted schematically in Figure 6, and described in greater detail below.

cDNA coding for the human GluR3A receptor was identified by probing human hippocampal cDNA that was obtained as an EcoRI-based lambda phage library (lambda ZAP) from Stratagene Cloning Systems (La Jolla, California, U.S.A.). The cDNA library was probed initially with a 1.1kb EcoRI/EcoRI DNA fragment constituting the 3' region of a kainate-binding human EAA receptor, designated humEAAla. This particular kainate-binding receptor is described in EP-A-0 529 994 incorporated herein by reference. DNA coding for the human EAA1a receptor, and from which the 1.1kb probe may be recovered, was deposited under terms of the Budapest Treaty, with the American Type Culture Collection in Rockville, Maryland U.S.A. on August 21, 1991 under accession number ATCC 75063.

Hybridizations using the probe were carried out at 30C overnight, and filters were washed with 2xSSC containing 0.5% SDS at 25C for 5 minutes, followed by a 15 minute wash at 50C with 2xSSC containing 0.5% SDS. The final wash was with 1xSSC containing 0.5% SDS at 50C for 15 minutes. Filters were exposed to X-ray film (Kodak) overnight. Of 10<sup>d</sup> clones screened under the following hybridization conditions (6xSSC, 50% formamide, 5% Denhardt's solution, 0.5% SDS, 100ug/ml denatured salmon sperm DNA), only two hippocampal cDNA library inserts were identified, one about 1.6kb and designated RKCH521 and another about 2.2kb and designated RKCH221 (Fig.6). For sequencing, the '521 and the '221 phages were plaque purified, then excised as phagemids according to the supplier's specifications, to generate insert-carrying Bluescript-SK variants of the phagemid vector. Sequencing of the '221 done across its entire sequence revealed a putative ATG

initiation codon together with about 78 bases of 5'non-coding region and about 2.1 kb of coding region. Sequencing across the '521 insert revealed a significant region of overlap with the '221 insert, and provided some additional 3' sequence, although no termination codon was located.

There being no termination codon apparent in the '521 sequence, a 3' region of the gene was sought. For this purpose, there was first synthesized an oligonucleotide probe capable of annealing to the 3' region of the rat GluR3 receptor sequence reported by Keinanen et al. supra. The specific sequence of the 32-P-labelled probe is provided below (SEQ ID NO: 13):

# 5'-ACACTCAGAATTACGCTACATACAGAGAAGGCTACAACGT-3'

The same hippocampal cDNA library was then re-screened using the rat-based probe and under the following hybridization conditions; 6xSSC, 25% formamide, 5% Dernhardt's solution, 0.5% SDS, 100ug/ml denatured salmon sperm DNA, 42C. This revealed a 1.2kb insert, designated RKCSHG132. Sequencing of the entire insert revealed 5' overlap with the 3'end of the previously isolated '521 insert, and also revealed a termination codon as well as about 15 bases of 3'non-translated sequence.

To provide the entire coding region in an intact clone, the strategy shown in Figure 6 was employed, to generate the phagemid pBS/HumGluR3A which carries the hGluR3A-encoding DNA as a 2.8kb EcoRI/EcoRI insert in a 3.0kb Bluescript-SK phagemid background. The entire sequence of the EcoRI/EcoRI insert is provided in Figure 3 (SEQ ID NOS: 5 and 6).

The 5.8kb phagemid pBS/humGluR3A was deposited, under the terms of the Budapest Treaty, with the American Type Culture Collection in Rockville, Maryland USA on March 19, 1992, and has been assigned accession number ATCC 75218.

#### Example 2 - Isolation of DNA coding for human GluR3B receptor

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A human fetal brain cDNA library was also screened in the search for human GluR receptors. Thisparticular library was obtained as an EcoRI-based lambda gt10 library from Strategene Cloning Systems (La Jolla, California, U.S.A.). The library was first screened using as hybridization probe an oligonucleotide capable of hybridizing to a 3' region of the reported rat GluR3 gene sequence. Screening using hybridization conditions as noted above (6xSSC, 25% formamide, 42C, etc.) revealed one insert about 2.3kb in size, designated RKCSFG34. After excision to release Bluescript-SK phagemids carrying the insert, sequencing revealed substantial sequence identity between the '34 insert and the 3'end of the earlier isolated GluR3A clone, and suggested that the 5'end of the gene encoded on partially on the '34 insert was missing. To provide an assembled gene, a 5' region was excised from the GluR3A insert and used to generate the 5'end of the '34 insert, at an internal HindIII site. This was achieved as depicted schematically in Figure 7. The resulting intact clone was designated human GluR3B.

Sequence comparison between the GluR3A clone of Example 1 and the GluR3B clone of this Example revealed only a short region of dissimilarity which is illustrated, in terms of amino acid sequence, in Figure 5 (SEQ ID NOS: 9 and 10).

The 6.1kb phagemid pBS/humGluR3B was deposited, under the terms of the Budapest Treaty, with the American Type Culture Collection in Rockville, Maryland USA on March 19, 1992, and has been assigned accession number ATCC 75219.

#### Example 3 - Isolation of DNA coding for the human GluR1B receptor

cDNA coding for the human GluR1B receptor was identified by probing human fetal brain cDNA that was obtained as an EcoRI-based lambda phage library (lambda ZAP) from Stratagene Cloning Systems (La Jolla, California, U.S.A.). The cDNA library was screened using an oligonucleotide probe capable of annealing to the 5' region of the rat GluR1 receptor sequence reported by Hollmann et al, supra. The specific sequence of the 32-P-labelled probe is provided below (SEQ ID NO: 14):

#### 5'-CCAGATCGATATTGTGAACATCAGCGACACGTTTGAGATG-3'

The fetal brain cDNA library was screened under the following hybridization conditions; 6xSSC, 25% formamide, 5% Dernhardt's solution, 0.5% SDS, 100ug/ml denatured salmon sperm DNA, 42C. Filters were washed with 2xSSC containing 0.5% SDS at 25C for 5 minutes, followed by a 15 minute wash at 50C with 2xSSC containing 0.5% SDS. The final wash was with 1xSSC containing 0.5% SDS at 50C for 15 minutes. Filters were exposed to X-ray film (Kodak) overnight. Of 10<sup>s</sup> clones screened, only one cDNA insert, of about 3.2kb, was

identified, and designated RKCSFG91. For sequencing, the '91 phage was plaque purified, then excised as a phagemid according to the supplier's specifications, to generate an insert-carrying Bluescript-SK variant of the phagemid vector. Sequencing of the '91 clone across its entire sequence revealed a putative ATG initiation codon together with about 61 bases of 5'non-coding region and 2,718 bases of coding region. Also revealed was a termination codon, as well as about 438 bases of 3' non-translated sequence. The entire sequence of the EcoRI/EcoRI insert is provided in Figure 1 (SEQ ID NOS: 1 and 2).

A 6.2kb phagemid designated pBS/humGluR1B, carrying the receptor-encoding DNA as a 3.2kb EcoRI/EcoRI insert in a 3.0kb Bluescript-SK phagemid background, was deposited, under the terms of the Budapest Treaty, with the American Type Culture Collection in Rockville, Maryland USA on May 28, 1992, and has been assigned accession number ATCC 75246.

#### Example 4 - Isolation of DNA coding for the human GluR2B receptor

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The particular strategy used to clone the human Glu2B receptor is depicted schematically in Figure 10, and described in greater detail below.

cDNA coding for the human GluR2B receptor was identified by probing human hippocampal cDNA that was obtained as an EcoRI-based lambda phage library (lambda ZAP) from Stratagene Cloning Systems (La Jolla, California, U.S.A.). The cDNA library was screened using an oligonucleotide probe capable of annealing to the 3' region of the rat GluR2 receptor sequence reported by Keinanen et al, supra. The specific sequence of the 32-P-labelled probe is provided below (SEQ ID NO: 15):

#### 5'-GTGAATGTGGAGCCAAGGACTCGGGAAGTAAG-3'

The hippocampal cDNA library was screened under the following hybridization conditions; 6xSSC, 25% formamide, 5% Dernhardt's solution, 0.5% SDS, 100ug/ml denatured salmon sperm DNA, 42C. Filters were washed with 2xSSC containing 0.5% SDS at 25C for 5 minutes, followed by a 15 minute wash at 50C with 2xSSC containing 0.5% SDS. The final wash was with 1xSSC containing 0.5% SDS at 50C for 15 minutes. Filters were exposed to X-ray film (Kodak) overnight. Of 10° clones screened, only two cDNA inserts were identified, one about 2.7kb and designated RKCSHG84 and another about 2.9kb and designated RKCSHG41 (Fig.10). For sequencing, the '84 and the '41 phages were plaque purified, then excised as phagemids according to the supplier's specifications, to generate insert-carrying Bluescript-SK variants of the phagemid vector. Sequencing of the '84 clone across its entire sequence revealed a putative ATG initiation codon together with about 314 bases of 5'non-coding region and about 2.4 kb of coding region. Sequencing across the '41 insert revealed a significant region of overlap with the '84 insert, and also revealed a termination codon not found in the '84 insert as well as about 441 bases of 3' non-translated sequence.

To provide the entire coding region in an intact clone, the strategy shown in Figure 10 was employed, to generate the phagemid pBS/HumGluR2B which carries the hGluR2B-encoding DNA as a 3.4kb EcoRI/Pstl insert in a 3.0kb Bluescript-SK phagemid background. The entire sequence of the EcoRI/Pstl insert is provided in Figure 2 (SEQ ID NOS: 3 and 4).

The 6.4kb phagemid pBS/humGluR2B was deposited, under the terms of the Budapest Treaty, with the American Type Culture Collection in Rockville, Maryland USA on March 19, 1992, and has been assigned accession number ATCC 75217.

# 5 Example 5 - Construction of genetically engineered cells producing human GluR3A receptors

The strategy depicted in Figure 8 was employed to facilitate incorporation of the GluR3A receptor-encoding cDNA into an expression vector.

For transient expression in mammalian cells, cDNA coding for the human GluR3A receptor was incorporated into the mammalian expression vector pcDNAI, which is available commercially from Invitrogen Corporation (San Diego, California, USA; catalogue number V490-20). This is a multifunctional 4.2kb plasmid vector designed for cDNA expression in eukaryotic systems, and cDNA analysis in prokaryotes. Incorporated on the vector are the CMV promoter and enhancer, splice segment and polyadenylation signal, an SV40 and Polyoma virus origin of replication, and M13 origin to rescue single strand DNA for sequencing and mutagenesis, Sp6 and T7 RNA promoters for the production of sense and anti-sense RNA transcripts and a Col E1-like high copy plasmid origin. A polylinker is located appropriately downstream of the CMV promoter (and 3' of the T7 promoter).

To facilitate incorporation of the GluR3A receptor-encoding cDNA into an expression vector, a Notl site was introduced onto the 5' flank of the Bluescript-SK cDNA insert, and the cDNA insert was then released from

pBS/humGluR3A as a 2.8 kb Notl/Notl fragment, which was then incorporated at the Notl site in the pcDNAI polylinker. Sequencing across the Notl junction was performed, to confirm proper insert orientation in pcDNAI. The resulting plasmid, designated pcDNAI/humGluR3A, was then introduced for transient expression into a selected mammalian cell host, in this case the monkey-derived, fibroblast like cells of the COS-1 lineage (available from the American Type Culture Collection, Rockville, Maryland as ATCC CRL 1650).

For transient expression of the GluR3A-encoding DNA, COS-1 cells were transfected with approximately 8ug DNA (as pcDNA1/humGluR3A) per 10<sup>6</sup> COS cells, by DEAE-mediated DNA transfection and treated with chloroquine according to the procedures described by Maniatis et al, supra. Briefly, COS-1 cells were plated at a density of 5 x 10<sup>6</sup> cells/dish and then grown for 24 hours in FBS-supplemented DMEM/F12 medium. Medium was then removed and cells were washed in PBS and then in medium. There was then applied on the cells 10ml of a transfection solution containing DEAE dextran (0.4mg/ml), 100uM chloroquine, 10% NuSerum, DNA (0.4mg/ml) in DMEM/F12 medium. After incubation for 3 hours at 37C, cells were washed in PBS and medium as just described and then shocked for 1 minute with 10% DMSO in DMEM/F12 medium. Cells were allowed to grow for 2-3 days in 10% FBS-supplemented medium, and at the end of incubation dishes were placed on ice, washed with ice cold PBS and then removed by scraping. Cells were then harvested by centrifugation at 1000 rpm for 10 minutes and the cellular pellet was frozen in liquid nitrogen, for subsequent use in ligand binding assays. Northern blot analysis of a thawed aliquot of frozen cells confirmed expression of receptor-encoding cDNA in cells under storage.

In a like manner, stably transfected cell lines can also prepared using two different cell types as host: CHO K1 and CHO Pro5. To construct these cell lines, cDNA coding for human GluR3A was incorporated into the mammalian expression vector pRC/CMV (Invitrogen), which enables stable expression. Insertion at this site placed the cDNA under the expression control of the cytomegalovirus promoter and upstream of the polyadenylation site and terminator of the bovine growth hormone gene, and into a vector background comprising the neomycin resistance gene (driven by the SV40 early promoter) as selectable marker.

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To introduce plasmids constructed as described above, the host CHO cells are first seeded at a density of 5 x 10<sup>5</sup> in 10% FBS-supplemented MEM medium. After growth for 24 hours, fresh medium are added to the plates and three hours later, the cells are transfected using the calcium phosphate-DNA co-precipitation procedure (Maniatis et al, supra). Briefly, 3ug of DNA is mixed and incubated with buffered calcium solution for 10 minutes at room temperature. An equal volume of buffered phosphate solution is added and the suspension is incubated for 15 minutes at room temperature. Next, the incubated suspension is applied to the cells for 4 hours, removed and cells were shocked with medium containing 15% glycerol. Three minutes later, cells are washed with medium and incubated for 24 hours at normal growth conditions. Cells resistant to neomycin are selected in 10% FBS-supplemented alpha-MEM medium containing G418 (1mg/ml). Individual colonies of G418-resistant cells are isolated about 2-3 weeks later, clonally selected and then propogated for assay purposes.

# Example 6 - Construction of genetically engineered cells producing human GluR1B receptors

The strategy depicted in Figure 9 was employed to facilitate incorporation of the GluR1B receptor-encoding cDNA into an expression vector. Particularly, a NotI site was introduced onto the 3' flank of the Bluescript-SK cDNA insert, and the cDNA insert was then released from pBS/humGluR1B as a 3.2kb NotI/NotI fragment, which was then incorporated at the NotI site in the pcDNAI polylinker. Sequencing across the junctions was performed, to confirm proper insert orientation in pcDNA1. The resulting plasmid, designated pcDNA1/ hum-GluR1B, was then introduced for transient expression into monkey-derived, fibroblast like cells of the COS-1 lineage as described above.

For transient expression of the GluR1B-encoding DNA, COS-1 cells were transfected with approximately 8ug DNA (as pcDNA1/humGluR1B) per 10<sup>6</sup> COS cells using the method described in Example 5.

#### Example 7 - Construction of genetically engineered cells producing human GluR2B receptors

The strategy depicted in Figure 11 was employed to facilitate incorporation of the GluR2B receptor-encoding cDNA into an expression vector. Particularly, a NotI site was introduced onto the 5' flank of the Bluescript-SK cDNA insert, and the cDNA insert was then released from pBS/humGluR2B as a 3.4kb HindIII/NotI fragment, which was then incorporated at the HindIII/NotI sites in the pcDNAI polylinker. Sequencing across the junctions was performed, to confirm proper insert orientation in pcDNA1. The resulting plasmid, designated pcDNA1/humGluR2B, was then introduced for transient expression into a selected mammalian cell host, in this case the monkey-derived, fibroblast like cells of the COS-1 lineage (available from the American Type Culture Collection, Rockville, Maryland as ATCC CRL 1650).

For transient expression of the GluR2B-encoding DNA, COS-1 cells were transfected with approximately 8ug DNA (as pcDNA1/humGluR2B) per 10<sup>8</sup> COS cells as set out in Example 5.

#### Example 8 - Ligand binding assays

Transfected cells in the frozen state were resuspended in ice-cold distilled water using a hand homogenizer, sonicated for 5 seconds, and then centrifuged for 20 minutes at 50,000g. The supernatant was discarded and the membrane pellet stored frozen at -70C.

COS cell membrane pellets were suspended in ice cold 50mM Tris-HCl (pH 7.55, 5C) and centrifuged again at 50,000g for 10 minutes in order to remove endogenous glutamate that would compete for binding. Pellets were resuspended in ice cold 50mM Tris-HCl (pH 7.55) buffer and the resultant membrane preparation was used as tissue source for binding experiments described below. Proteins were determined using the Pierce Reagent with BSA as standard.

Binding assays were then performed, using an amount of COS-derived membrane equivalent to from 25-100ug as judged by protein determination and selected radiolabelled ligand. In particular, for AMPA-binding assays, incubation mixtures consisted of 25-100ug tissue protein and D,L-alpha-[5-methyl-3H]amino-3-hydroxy-5-methylisoxazole-4-propionic acid (3H-AMPA, 27.6Ci/mmole, 10nM final) with 0.1M KSCN and 2.5mM CaCl<sub>2</sub> in the 1ml final volume. Non-specific binding was determined in the presence of 1mM L-glutamate. Samples were incubated on ice for 60 minutes in plastic minivials, and bound and free ligand was separated by centrifugation for 30 minutes at 50,000g. Pellets were washed twice in 6ml of the cold incubation buffer, then 5ml of Beckman Ready-Protein Plus scintillation cocktail was added, for counting.

For kainate-binding assays, incubation mixtures consisted of 25-100ug tissue protein and [vinylidene-3H] kainic acid (58Ci/mmole, 5nM final) in the cold incubation buffer, 1ml final volume. Non-specific binding was determined in the presence of 1mM L-glutamate. Samples were incubated as for the AMPA-binding assays, and bound and free ligand were separated by rapid filtration using a Brandel cell harvester and GF/B filters pre-soaked in ice-cold 0.3% polyethyleneimine. Filters were washed twice in 6ml of the cold incubation buffer, then placed in scintillation vials with 5ml of Beckman Ready-Protein Plus scintillation cocktail for counting.

Assays performed in this manner, using membrane preparations derived from the human GluR3A receptor-producing COS cells, revealed specific binding of 25-30 fmole/mg protein at 10nM [³H]-AMPA (Figure 14); using membrane preparations derived from the human GluR1B receptor-producing COS cells, specific binding of about 100-150 fmole/mg protein at 10nM [³H]-AMPA was revealed (Figure 12); and using membrane preparations derived from the human GluR2B receptor-producing COS cells, specific binding of 750-850 fmol/mg protein at 10nM [³H]-AMPA was revealed (Figure 13). Mock transfected cells exhibited no specific binding of any of the linands tested

Scatchard analysis indicated that the recombinantly expressed human GluR1B and GluR2B receptors each contain a single class of [ $^3$ H]-labelled AMPA binding sites with a dissociation constants (Kd) of 46 nM (Figure 15) and about 36.3  $\pm$  7.4 nM (Figure 16), respectively. Further, the maximum AMPA-binding of the GluR1B and GluR2B receptors has been found to be 847 and 816  $\pm$  302 fmol/mg protein, respectively.

[3H]-AMPA displacement assays have also been performed for the GluR2B receptors in COS cells to determine the relative binding affinity of selected ligands. These results, as illustrated in Figure 17, indicate the rank order of potency of the ligands in displacing <sup>3</sup>H-AMPA binding to the GluR2B receptor to be as follows:

quisqualate = AMPA > DNQX > CNQX > glutamate > domoate > kainate

These results demonstrate clearly that the human GluR receptors bind AMPA with specificity. This activity, coupled with the fact that there is little or no demonstrable binding of either kainate or NMDA, clearly assigns the human GluR receptors to be of the AMPA type of EAA receptor. Furthermore, this binding profile indicates that the receptor is binding in an authentic manner, and can therefore reliably predict the ligand binding "signature" of its non-recombinant counterpart from the human brain. These features make the recombinant receptor especially useful for selecting and characterizing ligand compounds which bind to the receptor, and/or for selecting and characterizing compounds which may act by displacing other ligands from the receptor. The isolation of the GluR receptor genes in substantially pure form, capable of being expressed as a single, homogeneous receptor species, therefore frees the ligand binding assay from the lack of precision introduced when complex, heterogeneous receptor preparations from human and other mammalian brains are used to attempt such characterizations.

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# SEQUENCE LISTING

5	(1) GF	ENERAL INFORMATION:
10		(i) APPLICANT:  (A) NAME: ALLELIX BIOPHARMACEUTICALS  (B) STREET: 6850 Goreway Drive  (C) CITY: Mississauga  (D) STATE OR PROVINCE: Ontario  (E) COUNTRY: Canada  (F) POSTAL CODE: L4V 1P1  (G) TELEPHONE: (416) 677-0831  (H) FAX: (416) 677-9595
15	i)	i) title of invention: AMPA-BINDING HUMAN GLUTAMATE RECEPTORS
	(ii	i) NUMBER OF SEQUENCES: 15
20	(i	LV) COMPUTER READABLE FORM:  (A) MEDIUM TYPE: Floppy disk  (B) COMPUTER: IBM FC compatible  (C) OPERATING SYSTEM: PC-DOS/MS-DOS  (D) SOFTWARE: Patentin Release \$1.0, Version \$1.25
	(	v) CURRENT APPLICATION DATA: (A) APPLICATION NUMBER: Unknown
25	(1	(A) APPLICATION DATA:  (A) APPLICATION NUMBER: US 07/896,437  (B) FILING DATE: 10-JUN-1992
	(٧	(A) APPLICATION DATA:  (B) FILING DATE: 10-JUN-1992
		(A) APPLICATION DATA:  (B) FILING DATE: 10-JUN-1992
35	(2) IN	FORMATION FOR SEQ ID NO:1:
	(	i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 3220 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: double  (D) TOPOLOGY: linear
40	(i	i) MOLECULE TYPE: cDNA
45	i)	(A) NAME/KEY: CDS (B) LOCATION: 622782
	(i	(A) NAME/KEY: sig_peptide (B) LOCATION: 62115
50	į)	x) FEATURE: (A) NAME/KEY: mat_peptide

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15	CCA Pro	AAC Asn 15	CAG Gln	CAG Gln	TCA Ser	CAG Gln	GAA Glu 20	CAT His	GCT Ala	GCT Ala	TTT Phe	AGA Arg 25	TTT Phe	GCT Ala	TTG Leu	TCG Ser	202	2
. •	CAA Gln 30	CTC Leu	ACA Thr	GAG Glu	CCC	CCG Pro 35	AAG Lys	CTG Leu	CTC Leu	CCC Pro	CAG Gln 40	Ile	GAT Asp	ATT Ile	GTG Val	AAC Aen 45	250	)
20	ATC Ile	AGC Ser	GAC Asp	ACG Thr	TTT Phe 50	GAG Glu	ATG Met	ACC Thr	TAT Tyr	AGA Arg 55	TTC Phe	TGT Cys	TCC Ser	CAG Gln	TTC Phe 60	TCC Ser	298	3 _
25	Lys	GGA Gly	GTC Val	TAT Tyr 65	GCC Ala	ATC Ile	TTT Phe	GGG Gly	TTT Phe 70	TAT Tyr	GAA Glu	CGT Arg	AGG Arg	ACT Thr 75	GTC Val	AAC Asn	340	5
	ATG Met	CTG Leu	ACC Thr 80	TCC Ser	TTT Phe	TGT Cys	GGG Gly	GCC Ala 85	CTC Leu	CAC His	GTC Val	TGC Cyb	TTC Phe 90	ATT Ile	ACG Thr	CCG Pro	394	ŧ
30	AGC Ser	TTT Phe 95	CCC Pro	GTT Val	GAT Asp	ACA Thr	TCC Ser 100	AAT Asn	CAG Gln	TTT Phe	GTC Val	CTT Leu 105	CAG Gln	CTG Leu	CGC Arg	CCT Pro	44:	2
25	GAA Glu 110	CTG Leu	CAG Gln	GAT Asp	GCC Ala	CTC Leu 115	ATC Ile	AGC Ser	ATC Ile	ATT Ile	GAC Asp 120	CAT His	TAC Tyr	AAG Lys	TGG Trp	CAG Gln 125	490	כ
35	FÅR	TTT Phe	GTC Val	TAC Tyr	ATT Ile 130	TAT Tyr	GAT Asp	GCC Ala	GAC <b>As</b> p	CGG Arg 135	GGC Gly	TTA Leu	TCC Ser	GTC Val	CTG Leu 140	CAG Gln	53,	3
40	AAA Lys	GTC Val	CTG Leu	GAT Asp 145	ACA Thr	GCT Ala	GCT Ala	GAG Glu	AAG Lys 150	<b>As</b> n	TGG Trp	CAG Gln	GTG Val	ACA Thr 155	GCA Ala	GTC Val	58	5
	AAC Asn	ATT Ile	TTG Leu 160	ACA Thr	ACC Thr	ACA Thr	GAG Glu	GAG Glu 165	GGA Gly	TAC Tyr	CGG Arg	ATG Met	CTC Leu 170	TTT Phe	CAG Gln	GAC Asp	63	4
45	CTG Leu	GAG Glu 175	AAG Lyb	AAA Lys	AAG Lys	Glu	CGG Arg 180	CTG Leu	GTG Val	GTG Val	GTG Val	GAC Asp 185	тст Сув	GAA Glu	TCA Ser	GAA Glu	68:	2
50	CGC Arg 190	CTC Leu	AAT Asn	GCT Ala	ATC Ile	TTG Leu 195	GCC Gly	CAG Gln	ATT Ile	ATA Ile	AAG Lys 200	CTA Leu	GAG Glu	AAG Lyb	AAT Asn	GGC Gly 205	73	0
	ATC Ile	GLY	TAC Tyr	CAC His	TAC Tyr 210	ATT Ile	CTT Leu	GCA Ala	AAT Asn	CTG Leu 215	GGC Gly	TTC Phe	ATG Met	GAC Asp	ATT Ile 220	GAC Asp	77	3
55	TTA Leu	AAC Asn	AAA Lys	TTC Phe	AAG Lýs	GAG Glu	AGT Ser	GGC Gly	GCC Ala	AAT Asn	GTG Val	ACA Thr	GGT Gly	TTC Phe	CAG Gln	CTG Leu	82	5

5	GTG Val	AAC Asn	TAC Tyr 240	ACA Thr	GAC Asp	ACT Thr	ATT Ile	CCG Pro 245	GCC Ala	AAG Lys	ATC Ile	ATG Ket	CAG Gln 250	CAG Gln	TGG Trp	AAG Lys		874
	AAT Asn	AGT Ser 255	GAT Asp	GCT Ala	CGA Arg	GAC Asp	CAC His 260	ACA Thr	CGG Arg	GTG Val	GAC Asp	TGG Trp 265	ÀÀĞ Lys	AGA Arg	CCC Pro	AAG Lys		922
10	TAC Tyr 270	ACC Thr	TCT Ser	GCG Ala	CTC Leu	ACC Thr 275	TAC Tyr	GAT Asp	GGG Gly	GTG Val	AAG Lys 280	GTG Val	ATG Met	GCT Ala	GAG Glu	GCT Ala 285		970
15	TTC Phe	CAG Gln	AGC Ser	CTG Leu	CGG Arg 290	AGG Arg	CAG Gln	AGA Arg	ATT Ile	GAT Asp 295	ATA Ile	TCT Ser	CGC Arg	CGG Arg	GGG Gly 300	AAT Asn	1	.018
	GCT Ala	GGG Gly	GAT Asp	TGT Cys 305	Leu	GCT Ala	AAC Asn	CCA Pro	GCT Ala 310	GTT Val	CCC Pro	TGG Trp	età ecc	CAA Gln 315	GGG Gly	ATC Ile	1	.066
20	GAC Asp	ATC Ile	CAG Gln 320	AGA Arg	GCT Ala	CTG Leu	CAG Gln	CAG Gln 325	GTG Val	CGA Arg	TTT Phe	GAA Glu	GCT Gly 330	TTA Leu	ACA Thr	GGA Gly	1	114
25	AAC Asn	GTG Val 335	CAG Gln	TTT Phe	AAT Asn	GAG Glu	AAA Lys 340	GGA Gly	CGC Arg	CGG Arg	ACC Thr	AAC Asn 345	TAC Tyr	ACG Thr	CTC Leu	CAC His	1	162
	GTG Val 350	ATT Ile	GAA Glu	ATG Met	AAA Lys	CAT His 355	GAC Asp	GGC Gly	ATC Ile	CGA Arg	AAG Lys 360	ATT Ile	GGT Gly	TAC Tyr	TGG Trp	AAT Asn 365	1	210
30	GAA Glu	GAT Asp	gat Asp	AAG Lys	TTT Phe 370	GTC Val	CCT Pro	GCA Ala	GCC Ala	ACC Thr 375	GAT Asp	GCC Ala	CAA Gln	GCT Ala	GGG Gly 380	GGC Gly	1	.258
	GAT Asp	AAT Asn	TCA Ser	AGT Ser 385	GTT Val	CAG Gln	AAC Asn	AGA Arg	ACA Thr 390	TAC Tyr	ATC Ile	GTC Val	ACA Thr	ACA Thr 395	ATC Ile	CTA Leu	1	.306
35	GAA Glu	GAT Asp	CCT Pro 400	TAT Tyr	GTG Val	ATG Met	CTC Leu	AAG Lys 405	AAG Lys	AAC Asn	GCC Ala	TAA Aan	CAG Gln 410	TTT Phe	GAG Glu	GCC	1	1354
40	AAT Asn	GAC Asp 415	CGT Arg	TAC Tyr	GAG Glu	GGC Gly	TAC Tyr 420	TGT Cys	GTA Val	GAG Glu	CTG Leu	GCG Ala 425	GCA Ala	GAG Glu	ATT Ile	GCC Ala	1	402
	AAG Lys 430	CAC His	GTG Val	GCC Gly	TAC Tyr	TCC Ser 435	TAC Tyr	CGT Arg	CTG Leu	GAG Glu	ATT Ile 440	GTC Val	AGT Ser	GAT Asp	GGA Gly	AAA Lys 445	1	1450
45	TAC Tyr	GGA Gly	GCC Ala	CGA Arg	GAC Asp 450	CCT Pro	GAC Asp	ACG Thr	AAG Lys	GCC Ala 455	TGG Trp	AAT Aan	GC Gly	ATG Met	GTG Val 460	GGA Gly	1	1498
	GAG Glu	CTG Leu	GTC Val	TAT Tyr 465	GGA Gly	AGA Arg	GCA Ala	GAT Asp	GTG Val 470	GCT Ala	GTG Val	GCT Ala	CCC Pro	TTA Leu 475	ACT Thr	ATC Ile	1	1546
50 -	ACT Thr	TTG Leu	GTC Val 480	CGG Arg	GAA Glu	GAA Glu	GTT Val	ATA Ile 485	GAT Asp	TTC Phe	TCC Ser	AAA Lys	CCA Pro 490	TTT Phe	ATG Met	AGT Ser	1	1594
55	TTG Leu	GGG Gly 495	ATC Ile	TCC Ser	ATC Ile	ATG Met	ATT Ile 500	AAA Lys	AAA Lys	CCA Pro	CAG Gln	AAA Lys 505	TCC Ser	AAG Lys	CCG Pro	GGT Gly	. 1	642

5	GTC Val 510	TTC Phe	TCC Ser	TTC Phe	CTT Leu	GAT Asp 515	CCT Pro	TTG Leu	GCT Ala	TAT Tyr	GAG Glu 520	ATT Ile	TGG Trp	ATG Met	TGC Cys	ATT 110 525	1690
10	Val	TTT Phe	Ala	Tyr	Ile 530	Gly	Val	Ser	Val	Val 535	Leu	Phe	Leu	Val	Ser 540	Arg	1738
,~	Pne	AGT Ser	Pro	Tyr 545	Glu	Trp	His	Ser	Glu 550	Glu	Phe	Glu	Glu	Gly 555	Arg	Asp	1786
15	Gln	ACA Thr	Thr 560	Ser	yab	Gln	Ser	<b>As</b> n 565	Glu	Phe	Gly	Ile	Phe 570	Asn	Ser	Leu	1834
	Trp	TTC Phe 575	Ser	Leu	Gly	Ala	Phe 580	Ket	Gln	Gln	Gly	Сув 585	Asp	Ile	Ser	Pro	1882
20	Arg 590	TCC Ser	Leu	ser	Gly	Arg 595	Ile	Val	Gly	Gly	Val 600	Trp	Trp	Phe	Phe	Thr 605	1930
25	TTA Leu	ATC Ile	ATC Ile	ATC Ile	TCC Ser 610	TCA Ser	TAT Tyr	ACA Thr	GCC Ala	AAT Asn 615	CTG Leu	GCC Ala	GCC Ala	TTC Phe	CTG Leu 620	ACC Thr	1978
	GTG Val	GAG Glu	AGG Arg	ATG Met 625	GTG Val	TCT Ser	CCC Pro	ATT	GAG Glu 630	AGT Ser	GCA Ala	GAG Glu	GAC Asp	CTA Leu 635	GCG Ala	AAC Asn	2026
30	GAG Glu	ACA Thr	GAA Glu 640	ATT Ile	GCC Ala	TAC Tyr	GGG Gly	ACG Thr 645	CTG Leu	GAA Glu	GCA Ala	GGA Gly	TCT Ser 650	ACT Thr	AAG Lys	GAG Glu	2074
	Phe	TTC Phe 655	Arg	Arg	Ser	Lys	Ile 660	Ala	Val	Phe	Glu	Lys 665	Met	Trp	Thr	Tyr	2122
35	ATG Met 670	AAG Lys	TCA Ser	GCA Ala	GAG Glu	CCA Pro 675	TCA Ser	GTT Val	TTT Phe	GTG Val	CGG Arg 680	ACC Thr	ACA Thr	GAG Glu	GAG Glu	GGG Gly 685	2170
.40	ATG Met	ATT Ile	CGA Arg	GTG Val	AGG Arg 690	AAA Lys	TCC Ser	AAA Lys	GGC Gly	AAA Lys 695	TAT Tyr	GCC Ala	TAC Tyr	CTC Leu	CTG Leu 700	GAG Glu	2218
	TCC Ser	ACC Thr	ATG Met	AAT Asn 705	GAG Glu	TAC Tyr	ATT Ile	GAG Glu	CAG Gln 710	CGG	AAA Lys	CCC Pro	TGT Cys	GAC Asp 715	ACC Thr	ATG Met	2266
45	AAG Lys	GTG Val	GGA Gly 720	GGT Gly	AAC Asn	TTG Leu	GAT Asp	TCC Ser 725	Lys	GGC Gly	TAT Tyr	GGC Gly	ATT Ile 730	GCA Ala	ACA Thr	CCC Pro	2314
50	AAG Lys	GGG Gly 735	Ser	GCC Ala	CTG Leu	Arg	GGT Gly 740	CCC Pro	GTA Val	AAC Asn	CTA Leu	GCG Ala 745	GTT Val	TTG Leu	AAA Lys	CTC Leu	2362
,	AGT Ser 750	GAG Glu	CAA Gln	ejà eec	GTC Val	TTA Leu 755	GAC Asp	AAG Lys	CTG Leu	AAA Lys	AGC Ser 760	AAA Lys	TCC Trp	TGG Trp	TAC Tyr	GAT Asp 765	2410
55	AAA Lys	GGG Gly	GAA Glu	TGT Cys	GGA Gly 770	AGC Ser	AAG Lys	GAC Asp	TCC Ser	GGA Gly 775	AGT Ser	AAG Lys	GAC Asp	AAG Lys	ACA Thr 780	AGC Ser	2458

5	GCT Ala	CTG Leu	AGC Ser	CTC Leu 785	AGC Ser	AAT Asn	GTG Val	GCA Ala	GGC Gly 790	GTG Val	TTC Phe	TAC Tyr	ATC Ile	CTG Leu 795	ATC Ile	GGA Gly	2506
10	GGA Gly	CTT Leu	GGA Gly 800	CTA Leu	GCC Ala	ATG Het	CTG Leu	GTT Val 805	GCC Ala	TTA Leu	ATC Ile	GAG Glu	TTC Phe 810	TGC	TAC Tyr	AAA Lys	2554
10	TCC Ser	CGT Arg 815	AGT Ser	GAA Glu	TCC Ser	AAG Lys	CGG Arg 820	ATG Met	AAG Lys	GGT Gly	TTT Phe	TGT Cys 825	TTG Leu	ATC Ile	CCA Pro	CAG Gln	2602
15	CAA Gln 830	TCC Ser	ATC Ile	AAC Asn	GAA Glu	GCC Ala 835	ATA Ile	CGG Arg	ACA Thr	TCG Ser	ACC Thr 840	CTC Leu	CCC Pro	CGC Arg	AAC Asn	AGC Ser 845	2650
	GGG Gly	GCA Ala	GGA Gly	GCC Ala	AGC Ser 850	AGC Ser	GGC Gly	GGC Gly	AGT Ser	GGA Gly 855	GAG Glu	AAT Asn	GCT Gly	CGG Arg	GTG Val 860	GTC Val	2698
20	AGC Ser	CAT His	GAC Asp	TTC Phe 865	CCC Pro	AAG Lys	TCC Ser	ATG Met	CAA Gln 870	TCG Ser	ATT Ile	CCT Pro	CA8 LCC	ATG Met 875	AGC Ser	CAC His	2746
25	AGT Ser	TCA Ser	GGG Gly 880	ATG Met	CCC Pro	TTG Leu	GGA Gly	GCC Ala 885	ACG Thr	GGA Gly	TTG Leu	TAAC	TGGI	AGC 1	AGATO	GGAGAC	2799
	ccc	rtgg	GA C	CAGO	CTC	e GC	CTCC	CAGO	ccc	ATC	CAA	ACC	TTC	AGT (	CCA	AAAACA	2859
	ACAI	ACAAJ	AT A	(GAA)	CCC	A AC	CAC	CACCI	ACC	ACTO	CGA	CCAC	AAG	AAG (	GATG!	ATTCAA	2919
30	CAG	TTT	rcc i	GAA0	AATT	G A	\AAA(	CAT	TTC	CTGT	ccc	TTTT	CCT	rtt :	TGA:	CTTCT	2979
30	TTC	ACCC	TT 3	CTG	TTG	T A	\GTG!	AGGAT	C GA	<b>LAAA</b>	AATA	CACT	CTAC	CTG (	CART	AAGGGG	3039
	AGAG	TAAC	cc 1	GTC	TAATO	A A	ACCT	STGTO	TC	GAGA	GTA	GAG	CAC	rcc .	AACA	CTAATG	3099
	AGG	AAAC1	rgc <i>i</i>	CTG	TTT	T T	TAA!	TCAC	TTO	TTAG	TGT	GTC	TAG:	IGT (	STGC	AATTTT	3159
35	TTT	CTT	ACT I	LATA	CCA	G G1	TTG	CAGG	TC	ctt)	\GGC	CCTT	MCC.	rtc '	rccr	GAATT	3219
	С																3220
																•	
40	(2)	INF															
			(i) S	(B)		GTH:	900 mino	ami	Lno . a Ld	: acid:	3						
45		(	Li) 1	COLEC	CULE	TYPI	3: p	rote	Ĺn								
		()	(i) S	EQUI	NCE	DES	CRIP	rion:	: SEÇ	OI C	NO:2	2:					
	Met -18	Gln	His	Ile -15	Phe	Ala	Phe	Phe	Cys -10	Thr	Gly	Phe	Leu	Gly -5	Ala	Val	
50	Val	Gly	Ala 1	Asn	Phe	Pro	Asn 5	Asn	Ile	Gln	Ile	Gly 10	Gly	Leu	Phe	Pro	
	Asn 15	Gln	Gln	Ser	Gln	Glu 20	His	Ala	Ala	Phe	<b>Arg</b> 25	Phe	Ala	Leu	Ser	Gln 30	
55	LON	The	Glu	Pro	Bro	T.va	Táu	Leu	Dro	Gla	T10	A a m	T1-	V-1	1	T1-	

	Ser	Авр	Thr	Phe 50	Glu	Met	Thr	Tyr	Arg 55	Phe	Cys	Ser	Gln	Phe 60	Ser	Lys
5	Gly	Val	Tyr 65	Ala	Ile	Phe	Gly	Phe 70	Tyr	Glu	Arg	Arg	Thr 75	Val	Asn	Met
	Leu	Thr 80	Ser	Phe	Сув	Gly	Ala 85	Leu	His	Val	Сув	Phe 90	Ile	Thr	Pro	Ser
10	Phe 95	Pro	Val	Asp	Thr	Ser 100	Asn	Gln	Phe	Val	Leu 105	Gln	Leu	Arg	Pro	Glu 110
	Leu	Gln	Asp	Ala	Leu 115	Ile	Ser	Ile	Ile	Asp 120	His	Tyr	Гåа	Trp	Gln 125	Lys
15	Phe	Val	Tyr	Ile 130	Tyr	Asp	Ala	Asp	Arg 135	Gly	Leu	Ser	Val	Leu 140	Gln	Lys
	Val	Leu	Asp 145	Thr	Ala	Ala	Glu	Lys 150	Asn	Trp	Gln	Val	Thr 155	Ala	Val	Asn
20	Ile	Leu 160	Thr	Thr	Thr	Glu	Glu 165	Gly	Tyr	Arg	Met	Leu 170	Phe	Gln	Asp	Leu
	Glu 175	Lys	Lys	Lys	Glu	Arg 180	Leu	Val	Val	Val	Asp 185	Сув	Glu	Ser	Glu	Arg 190
25	Leu	Asn	Ala	Ile	Leu 195	Gly	Gln	Ile	Ile	Lys 200	Leu	Glu	Lys	Asn	Gly 205	Ile
	Gly	Tyr	His	Tyr 210	Ile	Leu	Ala	Asn	Leu 215	Gly	Phe	Met	Asp	Ile 220	Asp	Leu
30	Asn	Lys	Phe 225	Lys	Glu	Ser	Gly	Ala 230	Asn	.Val	Thr	Gly	Phe 235	Gln	Leu	Val
	Asn	Tyr 240	Thr	Asp	Thr	Ile	Pro 245	Ala	Lys	Ile	Met	Gln 250	Gln	Trp	Lys	Asn
35	Ser 255	Asp	Ala	Arg	Asp	His 260	Thr	Arg	Val	Asp	Trp 265	Lys	Arg	Pro	Lys	Tyr 270
	Thr	Ser	Ala	Leu	Thr 275	Tyr	Asp	Gly	Val	Lys 280	Val	Met	Ala	Glu	Ala 285	Phe
40	Gln	Ser	Leu	Arg 290	Arg	Gln	Arg	Ile	Asp 295	Ile	Ser	Arg	Arg	Gly 300	Asn	Ala
	Gly	Asp	Сув 305	Leu	Ala	Asn	Pro	Ala 310	Val	Pro	Trp	Gly	Gln 315	Gly	Ile	Asp
45	Ile	Gln 320	Arg	Ala	Leu	Gln	Gln 325	Val	Arg	Phe	Glu	Gly 330	Leu	Thr	Gly	Asn
	Val 335	Gln	Phe	Asn	Glu	Lys 340	Gly	Arg	Arg	Thr	Asn 345	Tyr	Thr	Leu	His	Val 350
<b>50</b>	Ile	Glu	Met	Lys	His 355	Asp	ĆĮĀ	Ile	Arg	360 Lys	Ile	Gly	Tyr	Trp	Asn 365	Glu
	Asp	Asp	Lys	Phe 370	Val	Pro	Ala	Ala	Thr 375	qaA	Ala	Gln	Ala	Gly 380	Gly	Asp
55	Asn	Ser	Ser 385		Gln	Asn	Хrg	Thr 390		Ile	Val	Thr	Thr 395	Ile	Leu	Glu

	Asp	Pro 400	Tyr	Val	Met	Leu	Lys 405	Lys	Asn	Ala	Asn	Gln 410	Phe	Glu	Gly	Asn
5	Asp 415	Arg	Tyr	Glu	Gly	Tyr 420	Сув	Val	Glu	Leu	Ala 425	Ala	Glu	Île	Ala	Lys 430
	His	Val	Gly	Tyr	Ser 435	Tyr	Arg	Leu	Glu	Ile 440	Val	Ser	Авр	Gly	Lys 445	Tyr
10	Gly	Ala	Arg	Asp 450	Pro	Asp	Thr	Lys	Ala 455	Trp	Asn	Gly	Met	Val 460	Gly	Glu
	Leu	Val	Tyr 465	Gly	Arg	Ala	qaA	Val 470	Ala	Val	Ala	Pro	Leu 475	Thr	Ile	Thr
15	Leu	Val 480	Arg	Glu	Glu	Val	Ile 485	Asp	Phe	Ser	Lys	Pro 490	Phe	Met	Ser	Leu
	Gly 495	Ile	Ser	Ile	Met	Ile 500	Lys	Lys	Pro	Gln	Lys 505	Ser	Lys	Pro	Gly	Val 510
20 .	Phe	Ser	Phe	Leu	Asp 515	Pro	Leu	Ala	Tyr	Glu 520	Ile	Trp	Xet	Cys	Ile 525	Val
	Phe	Ala	Tyr	11e 530	Gly	Val	Ser	Val	Val 535	Leu	Phe	Leu	Val	Ser 540	Arg	Phe
25	Ser	Pro	Tyr 545	Glu	Trp	His	Ser	Glu 550	Glu	Phe	Glu	Glu	Gly 555	Arg	Asp	Gln
	Thr	Thr 560	Ser	yab	Gln	Ser	Asn 565	Glu	Phe	Gly	Ile	Phe 570	Asn	Ser	Leu	Trp
30	Phe 575	Ser	Leu	Gly	Ala	Phe 580	Met	Gln	Gln	Gly	Сув 585	Asp	Ile	Ser	Pro	Arg 590
	Ser	Leu	Ser	Gly	Arg 595	Ile	Val	Gly	Gly	Val 600	Trp	Trp	Phe	Phe	Thr 605	Leu
35	Ile	Ile	Ile	Ser 610	Ser	Tyr	Thr	Ala	Asn 615	Leu	Ala	Ala	Phe	Leu 620	Thr	Val
	Glu	Arg	Met 625	Val	Ser	Pro	Ile	Glu 630	Ser	Ala	Glu	Asp	Leu 635		Asn	Glu
40	Thr	Glu 640	Ile	Ala	Tyr	Gly	Thr 645	Leu	Glu	Ala	Gly	Ser 650	Thr	Lys	Glu	Phe
•	Phe 655	Arg	Arg	Ser	Lys	Ile 660	Ala	Val	Phe	Glu	Ly <b>s</b> 665	Ket	Trp	Thr	Tyr	Met 670
45	Lys	Ser	Ala	Glu	Pro 675	Ser	Val	Phe	Val	Arg 680	Thr	Thr	Glu	Glu	Gly 685	Met
	Ile	Arg	Val	Arg 690	Lys	Ser	Lys	Gly	Lys 695	Tyr	Ala	Tyr	Leu	Leu 700	Glu	Ser
50	Thr	Met	Asn 705	Glu	Tyr	Ile	Glu	Gln 710	Arg	Lys	Pro	САВ	Авр 715	Thr	Met	Lys
	Val	Gly 720	Gly	Asn	Leu	Asp	Ser 725	Lys	Gly	Tyr	Gly	11e 730	Ala	Thr	Pro	Lys
55	Gly 735	Ser	Ala	Leu	Arg	Gly 740	Pro	Val	Asn	Leu	Ala 745	Val	Leu	Lys	Leu	Ser 750

5	GIU	GIII	GLY	Val	755	veh	Lys	rad	ГÀВ	760	Lys	trb	rrp	TYF	765	гля	
	Gly	Glu	Сув	Gly 770	Ser	Lys	Asp	Ser	Gly 775	Ser	Lys	yab	Lys	Thr 780	Ser	Ala	
10	Leu	Ser	Leu 785	Ser	Asn	Val	Ala	Gly 790	Val	Phe	Tyr	Ile	Leu 795	Ile	Gly	Gly	
	Leu	Gly 800	Leu	Ala	Met	Leu	Val 805	Ala	Leu	Ile	Glu	Phe 810	Cys	Tyr	Lys	Ser	
	Arg 815	Ser	Glu	Ser	Lys	Arg 820	Met	Lys	Gly	Phe	Cys 825	Leu	Ile	Pro	Gln	Gln 830	
15	Ser	Ile	Asn	Glu	Ala 835	Ile	Arg	Thr	Ser	Thr 840	Leu	Pro	Arg	Asn	Ser 845	Gly	
	Ala	Gly	Ala	Ser 850	Ser	Gly	Gly	Ser	Gly 855	Glu	Asn	Gly	Arg	Val 860	Val	Ser.	
20	His	Asp	Phe 865	Pro	Lys	Ser	Met	Gln 870	Ser	Ile	Pro	Сув	Met 875	Ser	His	Ser	
	Ser	Gly 880	Met	Pro	Leu	Gly	Ala 885	Thr	Gly	Leu							
25																	
	(2)	INF	ORMA'	TION	FOR	SEQ	ID I	K:08	:								
		(i	) SE	QUEN A) L						rs							
				B) T' C) S'													
30				D) T					pra								
		(ii	) MO	LECU	LE T	YPE:	cDN:	A.									
35		(ix	) FE.	ATUR A) N		KBY:	CDS										
			(	B) L	OCAT	ION:	315	29	66					,			
		(ix	) FE	ATUR A) N B) L	ame/	KEY:	sig	_pep	tide								
40			•	•		1011.	313	,	•								
		(ix	) FE (	ATUR A) N		KEY:	mat	beb	tide								
			. (	B) L	OCAT	ION:	375	29	66								
45		(xi	) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:3:						
	GAA	TTCC	GTG	agtg	CATG	GG A	CCGT	CCTG	A AT	ATTC	CGAG	ACA	CTGG	GAC	CACA	GCGGCA	60
	GCT	CCGC	TGA	AAAC	TGCA	TT C	AGCC	agtc	c TC	CGGA	CITC	TGG	AGCG	GGG	ACAG	GGCGCA	120
	GGG	CATC	AGC	AGCC	ACCA	GC A	GGAC	CTGG	G AA	ATAG	GGAT	TCI	TCTG	CCT	CCAC	TTCAGG	180
50	TTT	TAGC	AGC	TTGG	TGCT	'AA A	TIGO	TGTC	T CA	TAAA	GCAG	AGG	ATCI	TAAT	TTGC	AGAGGA	240
	AAA	CAGC	CAA	agaa	GGAA	GA G	GAGG	AAAA	g ga	AAAA	AAAA	GGG	GTAI	ATT	GTGG	ATGCTC	300
55	TAC	TTTT	CTT	GGAA	ATG Met	Gln	AAG Lys	ATT	ATG Het	CAT His	Ile	TCI Ser	GTC Val	CTC Leu	CTI Leu	TCT Ser	350

5	CCT Pro	GTT Val	TTA Leu	TGG Trp -5	GGA Gly	CTG Leu	ATT Ile	TTT Phe	GGT Gly 1	GTC Val	TCT Ser	TCT Ser	AAC Asn 5	AGC Ser	ATA Ile	CAG Gln		398
10	ATA Ile	GGG Gly 10	GGG Gly	CTA Leu	TTT Phe	CCT Pro	AGG Arg 15	GGC Gly	GCC Ala	GAT Asp	CAA Gln	GAA Glu 20	TAC Tyr	AGT Ser	GCA Ala	TTT Phe		446
	CGA Arg 25	GTA Val	GGG Gly	ATG Met	GTT Val	CAG Gln 30	TTT Phe	TCC Ser	ACT Thr	TCG Ser	GAG Glu 35	TTC Phe	AGA Arg	CTG Leu	ACA Thr	CCC Pro 40	,	494
15	CAC His	ATC Ile	GAC Asp	AAT Asn	TTG Leu 45	GAG Glu	GTG Val	GCA Ala	AAC Asn	AGC Ser 50	TTC Phe	GCA Ala	GTC Val	ACT Thr	AAT Asn 55	GCT Ala		542
· 20	TTC Phe	TGC Cys	TCC Ser	CAG Gln 60	TTT Phe	TCG Ser	AGA Arg	GGA Gly	GTC Val 65	TAT Tyr	GCT Ala	ATT Ile	TTT Phe	GGA Gly 70	TTT Phe	TAT Tyr		590
	GAC Asp	AAG Lys	AAG Lys 75	TCT Ser	GTA Val	AAT Asn	ACC Thr	ATC Ile 80	ACA Thr	TCA Ser	TTT Phe	TGC Cys	GGA Gly 85	ACA Thr	CTC	CAC His		638
25	GTC Val	TCC Ser 90	TTC Phe	ATC Ile	ACT Thr	CCC Pro	AGC Ser 95	TTC Phe	CCA Pro	ACA Thr	GAT Asp	GGC Gly 100	ACA Thr	CAT His	CCA Pro	TTT Phe		686
	GTC Val 105	ATT Ile	CAG Gln	ATG Met	AGA Arg	CCC Pro 110	GAC Asp	CTC Leu	AAA Lys	GGA Gly	GCT Ala 115	CTC Leu	CTT Leu	AGC Ser	TTG Leu	ATT Ile 120		734
30											CTC Leu							782
35	GGC Gly	TTA Leu	TCA Ser	ACA Thr 140	CTG Leu	CAA Gln	GCT Ala	GTG Val	CTG Leu 145	GAT Asp	TCT Ser	GCT Ala	GCT Ala	GAA Glu 150	AAG Lys	AAA Lys		830
	TGG Trp	CAA Gln	GTG Val 155	ACT Thr	GCT Ala	ATC Ile	AAT Asn	GTG Val 160	GGA Gly	AAC Asn	ATT	AAC Asn	AAT Asn 165	GAC Asp	AAG Lys	AAA Lys		878
40	GAT Asp	GAG Glu 170	ATG Met	TAC Tyr	CGA Arg	TCA Ser	CTT Leu 175	TTT	CAA Gln	GAT Asp	CTG Leu	GAG Glu 180	TTA Leu	YYY Lys	AAG Lys	GAA Glu		926
											λλλ Lys 195							974
45	GAC Asp	CAG Gln	GTT Val	ATT	ACC Thr 205	ATT	GGA Gly	AAA Lys	CAC His	GTT Val 210	AAA Lys	GGG Gly	TAC Tyr	CAC	TAC Tyr 215	ATC Ile		1022
50											CTA Leu				Gln			1070
	GGA Gly	GGT Gly	GCA Ala 235	AAT Asn	GTC Val	TCT	GGA Gly	TTT Phe 240	CAG Gln	ATA Ile	GTG Val	GAC Asp	TAT Tyr 245	GAT Asp	GAT Asp	TCG Ser		1118
55			Ser													GAA Glu	•	1166

5	TAC Tyr 265	CCT Pro	GGA Gly	GCT Ala	CAC His	ACA Thr 270	ACA Thr	ACA Thr	ATT Ile	AAG Lys	TAT Tyr 275	ACT Thr	TCT Ser	GCT Ala	CTG Leu	ACC Thr 280	1214
	TAT Tyr	GAT Asp	GCC Ala	GTT Val	CAA Gln 285	GTG Val	ATG Ket	ACT Thr	GAA Glu	GCC Ala 290	TTC Phe	CGC Arg	AAC Asn	CTA Leu	AGG Arg 295	AAG Lys	1262
10	CAA Gln	AGA Arg	ATT Ile	GAA Glu 300	ATC Ile	TCC Ser	CGA Arg	AGG Arg	GGG Gly 305	AAT	GCA Ala	GGA Gly	GAC Asp	TGT Cys 310	CTG Leu	GCA Ala	1310
15	AAC Asn	CCA Pro	GCA Ala 315	GTG Val	CCC Pro	TGG Trp	GGA Gly	CAA Gln 320	GGT Gly	GTA Val	GAA Glu	ATA Ile	GAA Glu 325	AGG Arg	GCC Ala	CTC Leu	1358
	AAA Lys	CAG Gln 330	GTT Val	CAG Gln	GTT Val	GAA Glu	GGT Gly 335	CTC Leu	TCA Ser	GGA Gly	AAT Asn	ATA Ile 340	AAG Lys	TTT Phe	GAC Asp	CAG Gln	1406
20	AAT Asn 345	GGA Gly	AAA Lys	AGA Arg	ATA Ile	AAC Asn 350	TAT Tyr	ACA Thr	ATT Ile	AAC	ATC Ile 355	ATG Met	GAG Glu	CTC Leu	AAA Lys	ACT Thr 360	1454_
25	AAT Asn	GGG Gly	CCC Pro	CGG Arg	AAG Lys 365	Ile	GGC Gly	TAC Tyr	TGG Trp	AGT Ser 370	GAA Glu	GTG Val	GAC Asp	AAA Lys	ATG Met 375	GTT Val	1502
23	GTT Val	ACC Thr	CTT Leu	ACT Thr 380	GAG Glu	CTC Leu	CCT Pro	TCT Ser	GGA Gly 385	AAT Asn	GAC Asp	ACC Thr	TCT Ser	GGG Gly 390	CTT Leu	GAG Glu	1550
30	AAT Asn	AAG Lys	ACT Thr 395	GTT Val	GTT Val	GTC Val	ACC Thr	ACA Thr 400	ATT Ile	TTG Lau	GAA Glu	TCT Ser	CCG Pro 405	TAT Tyr	GTT Val	ATG Met	1598
	ATG Met	AAG Lys 410	AAA Lys	AAT Asn	CAT His	GAA Glu	ATG Met 415	CTT Leu	GAA Glu	GGC Gly	AAT Asn	GAG Glu 420	CGC Arg	TAT Tyr	GAG Glu	GGC Gly	1646
35	TAC Tyr 425	TGT Cys	GTT Val	GAC Asp	CTG Leu	GCT Ala 430	GCA Ala	GAA Glu	ATC Ile	GCC Ala	AAA Lys 435	CAT His	TGT Cys	GGG Gly	TTC Phe	AAG Lys 440	1694
40	TAC Tyr	AAG Lys	TTG Leu	ACA Thr	ATT Ile 445	GTT Val	GGT Gly	GAT Asp	GGC Gly	AAG Lys 450	TAT Tyr	GGG Gly	GCC Ala	AGG Arg	GAT Asp 455	GCA Ala	1742
	GAC Asp	ACG Thr	AAA Lys	ATT Ile 460	TGG Trp	AAT Asn	GGG Gly	ATG Net	GTT Val 465	GGA Gly	GAA Glu	CTT Leu	GTA Val	TAT Tyr 470	GGG Gly	AAA Lys	1790
45	GCT Ala	GAT Asp	ATT Ile 475	GCA Ala	ATT Ile	GCT Ala	CCA Pro	TTA Leu 480	ACT Thr	ATT Ile	ACC Thr	CTT Leu	GTG Val 485	AGA Arg	GAA Glu	GAG Glu	1838
	GTG Val	ATT 11e 490	GAC Asp	TTC Phe	TCA Ser	AAG Lys	CCC Pro 495	TTC Phe	ATG Met	AGC Ser	CTC Leu	GGG Gly 500	ATA Ile	TCT Ser	ATC Ile	ATG Met	1886
50	ATC Ile 505	AAG Lys	AAG Lys	CCT Pro	CAG Gln	AAG Lys 510	TCC Ser	AAA Lys	CCA Pro	GGA Gly	GTG Val 515	TTT Phe	TCC Ser	TTT Phe	CTT Leu	GAT Asp 520	1934
55	CCT Pro	TTA Leu	GCC Ala	TAT Tyr	GAG Glu 525	ATC Ile	TGG Trp	ATG Met	TGC Cys	ATT Ile 530	GTT Val	TTT Phe	GCC Ala	TAC Tyr	ATT Ile 535	GGG Gly	1982

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5	V41	Jei	. Val	540	Leu	Pne	Lau	Val	Ser 545	Arg	Phe	Ser	CCC Pro	Tyr 550	Glu	Trp	2030
10	nio	1112	555	GIU	Pne	GIU	Asp	560	Arg	Glu	Thr	Gln	AGT Ser 565	Ser	Glu	Ser	2078
	ACT Thr	AAT Asn 570	GIU	TTT Phe	GGG Gly	ATT	TTT Phe 575	AAT Asn	AGT Ser	CTC Leu	TGG Trp	TTT Phe 580	TCC Ser	TTG Leu	GGT Gly	GCC Ala	2126
15	TTT Phe 585	ne c	CGG Arg	CAA Gln	GGA Gly	TGC Cys 590	GAT Asp	ATT	TCG Ser	CCA Pro	AGA Arg 595	TCC Ser	CTC Leu	TCT Ser	GGG Gly	CGC Arg 600	2174
		Val	GIY,	GIY	605	Trp	тгр	Pne	Phe	610	Leu	Ile	ATA Ile	Ile	Ser 615	Ser	2222
20	-1-			620	200	776	ALG.	rne	625	Inr	Val	CIU	AGG Arg	Met 630	Val	Ser	2270
25	CCC Pro	ATC Ile	GAA Glu 635	AGT Ser	GCT Ala	GAG Glu	GAT Asp	CTT Leu 640	TCT Ser	AAG Lys	CAA Gln	ACA Thr	GAA Glu 645	ATT Ile	GCT Ala	TAT Tyr	2318
	GGA Gly	ACA Thr 650	TTA Leu	GAC Asp	TCT Ser	GGC Gly	TCC Ser 655	ACT Thr	AAA Lys	GAG Glu	TTT Phe	TTC Phe 660	AGG Arg	AGA Arg	TCT Ser	AAA Lys	2366
30	665	nia	A 4 1	File	vsb	670	Met	Trp	Thr	Tyr	Ket 675	Arg	AGT Ser	Ala	Glu	Pro 680	2414
	361	401	FILE	Aat	685	ine	Thr	WTS	GIu	690 GIÀ	Val	Ala	AGA Arg	Val	Arg 695	Lys	2462
35	TCC Ser	AAA Lys	GGG Gly	AAA Lys 700	TAT Tyr	GCC Ala	TAC Tyr	TTG Leu	TTG Leu 705	GAG Glu	TCC Ser	ACG Thr	ATG Met	AAC Asn 710	GAG Glu	TAC Tyr	2510
40	ATT	GAG Glu	CAA Gln 715	AGG Arg	AAG Lys	CCT Pro	TGC Cys	GAC Asp 720	ACC Thr	ATG Met	AAA Lys	GTT Val	GGT Gly 725	GGA Gly	AAC Aan	CTG Leu	2558
	GAT Asp	TCC Ser 730	AAA Lys	GGC Gly	TAT Tyr	GGC Gly	ATC Ile 735	GCA Ala	ACA Thr	CCT	AAA Lys	GGA Gly 740	TCC Ser	TCA Ser	TTA Leu	GGA Gly	2606
45	ACC Thr 745	CCA Pro	GTA Val	AAT Asn	CTT Leu	GCA Ala 750	GTA Val	TTG Leu	AAA Lys	CTC Leu	AGT Ser 755	GAG Glu	CAA Gln	GGC Gly	GTC Val	TTA Leu 760	2654
	GAC Asp	AAG Lys	CTG Leu	AAA Lys	AAC Asn 765	AAA Lye	TGG Trp	TGG Trp	TAC Tyr	GAT Asp 770	AAA Lys	GGT Gly	GAA Glu	TGT Cys	GGA Gly 775	GCC Ala	2702
50	AAG Lys	GAC Asp	TCT Ser	GGA Gly 780	AGT Ser	AAG Lys	GAA Glu	AAG Lye	ACC Thr 785	AGT Ser	GCC Ala	CTC Leu	AGT Ser	CTG Leu 790	AGC Ser	AAC Asn	2750
55	GTT Val	GCT Ala	GGA Gly 795	GTA Val	TTC Phe	TAC Tyr	ATC Ile	CTT Leu 800	GTC Val	ejà eee	GCC	CTT Leu	GGT Gly 805	TTG Leu	GCA Ala	ATG Met	2798

5	Leu Val Ala Leu Ile Glu Phe Cys Tyr Lys Ser Arg Ala Glu Ala Lys 810 815 820	284
10	CGA ATG AAG GTG GCA AAG AAT GCA CAG AAT ATT AAC CCA TCT TCC TCG Arg Met Lys Val Ala Lys Asn Ala Gln Asn Ile Asn Pro Ser Ser 825 830 835 840	289
10	CAG ART TCA CAG ART TTT GCA ACT TAT ARG GAA GGT TAC ARC GTA TAT Gln Asn Ser Gln Asn Phe Ala Thr Tyr Lys Glu Gly Tyr Asn Val Tyr 845 855	294
15.	GGC ATC GAA AGT GTT AAA ATT TAGGGGATGA CCTTGAATGA TGCCATGAGG Gly Ile Glu Ser Val Lys Ile 860	299
	AACAAGGCAA GGCTGTCAAT TACAGGAAGT ACTGGAGAAA ATGGACGTGT TATGACTCCA	305
	GAATTTCCCA AAGCNGTGCA TGCTGTCCCT TACGTGAGTC CTGGCATGGG AATGAATGTC	311
20	AGTGTGACTG ATCTCTCGTG ATTGATAAGA ACCTTTTGAG TGCCTTACAC AATGGTTTTC	317
	TTGTGTGTTT ATTGTCAAAG TGGTGAGAGG CATCCAGTAT CTTGAAGACT, TTTCTTTCAG	323
•	CCAAGAATTC TTAAATATGT GGAGTTCATC TTGAATTGTA AGGAATGATT AATTAAAACA	329
25	CAACATCTTT TTCTACTCGA GTTACAGACA AAGCGTGGTG GACATGCACA GCTAACATGG	335
	AAGTACTATA ATTTACCTGA AGTCTTTGTA CAGACAACAA ACCTGTTTCT GCAG	340
	(2) INFORMATION FOR SEQ ID NO:4:	
30	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 883 amino acids  (B) TYPE: amino acid  (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: protein	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
	Met Gln Lys Ile Met His Ile Ser Val Leu Leu Ser Pro Val Leu Trp -20 -15 -5	
40	Gly Leu Ile Phe Gly Val Ser Ser Asn Ser Ile Gln Ile Gly Gly Leu 1 5 10	
	Phe Pro Arg Gly Ala Asp Gln Glu Tyr Ser Ala Phe Arg Val Gly Met 15 20 25	
45	Val Gln Phe Ser Thr Ser Glu Phe Arg Leu Thr Pro His Ile Asp Asn 30 35 40	
	Leu Glu Val Ala Asn Ser Phe Ala Val Thr Asn Ala Phe Cys Ser Gln 45 50 55 60	
50	Phe Ser Arg Gly Val Tyr Ala Ile Phe Gly Phe Tyr Asp Lys Lys Ser 65 70 75	
	Val Asn Thr Ile Thr Ser Phe Cys Gly Thr Leu His Val Ser Phe Ile 80 85 90	
<b>.</b> .	Thr Pro Ser Phe Pro Thr Asp Gly Thr His Pro Phe Val Ile Gln Met 95 100 105	,

	Arg	Pro 110	Asp	Leu	Lys	Gly	Ala 115	Leu	Leu	Ser	Leu	Ile 120	Glu	Tyr	Tyr	Gln
5	Trp 125	Asp	Lys	Phe	Ala	Tyr 130	Leu	Tyr	Asp	Ser	Авр 135	Arg	Gly	Leu	Ser	Thr 140
	Leu	Gln	Ala	Val	Leu 145	qaA	Ser	Ala	Ala	Glu 150	Lys	Lys	Trp	Gln	Val 155	Thr
	Ala	Ile	Asn	Val 160	Gly	Asn	Ile	Asn	Asn 165	Asp	Lys	Lys	Авр	Glu 170	Met	Tyr
	Arg	Ser	Leu 175	Phe	Gln	Asp	Leu	Glu 180	Leu	Lys	Lys	Glu	Arg 185	Arg	Val	Ile
15	Leu	Авр 190	Cys	Glu	Arg	Asp	Lys 195	Val	Asn	Asp	Ile	Val 200	qsA	Gln	Val	Ile
	Thr 205	Ile	Gly	Lys	His	Val 210	Lys	Gly	Tyr	His	Tyr 215	Ile	Ile	Ala	Asn	Leu 220
	Gly	Phe	Thr	Asp	Gly 225	Asp	Leu	Leu	Lys	Ile 230	Gln	Phe	Gly	Gly	Ala 235	Asn
	Val	Ser	Gly	Phe 240	Gln	Ile	Val	Авр	Tyr 245	Asp	Asp	Ser	Leu	Val 250	Ser	Lys
25	Phe	Ile	Glu 255	Arg	Trp	Ser	Thr	Leu 260	Glu	Glu	Lys	Glu	Tyr 265	Pro	Gly	Ala
	His	Thr 270	Thr	Thr	Ile	ГÀв	Tyr 275	Thr	Ser	Ala	Leu	Thr 280	Tyr	Asp	Ala	Val
30	Gln 285	Val	Met	Thr	Glu	Ala 290	Phe	Arg	Asn	Leu	Arg 295	Lys	Gln	Arg	Ile	Glu 300
	Ile	Ser	Arg	Àrg	Gly 305	Asn	Ala	Gly	Asp	Cys 310	Leu	Ala	Asn	Pro	Ala 315	Val
35	Pro	Trp	Gly	Gln 320	Gly	Val	Glu	Ile	Glu 325	Arg	Ala	Leu	Lys	Gln 330	Val	Gln
	Val	Glu	Gly 335	Leu	Ser	Gly	Asn	Ile 340	Lys	Phe	Asp	Gln	Asn 345	Gly	Lys	Arg
<b>40</b>	Ile	Asn 350	Tyr	Thr	Ile	Asn	Ile 355	Met	Glu	Leu	Lys	Thr 360	Asn	Gly	Pro	Arg
,	Lys 365	Ile	Gly	Tyr	Trp	Ser 370	Glu	Val	Asp	Lys	Met 375	Val	Val	Thr	Leu	Thr 380
45	Glu	Leu	Pro	Ser	Gly 385	Asn	Авр	Thr	Ser	Gly Gly	Leu	Glu	Asn	Lys	Thr 395	Val
	Val	Val	Thr	Thr 400	Ile	Leu	Glu	Ser	Pro 405	Tyr	Val	Met	Met	Lys 410	Lys	Asn
,50	His	Glu	Met 415	Leu	Glu	Gly	Asn	Glu 420	Arg	Tyr	Glu	Gly	Tyr 425	Сув	Val	Asp
	Leu	Ala 430	Ala	Glu	Ile	Ala	Lys 435	His	Сув	Gly	Phe	Lys 440	Tyr	Lys	Leu	Thr
55	Ile 445	Val	Gly	Asp	Ġly	Lys 450	Tyr	Gly	Ala	Arg	Asp 455	Ala	Asp	Thr	Lys	Ile 460

	Trp	Asn	Gly	Met	Val 465	Gly	Glu	Leu	Val	Tyr 470	Gly	Lys	Ala	Авр	Ile 475	Ala
5	Ile	Ala	Pro	Leu 480	Thr	Ile	Thr	Leu	Val 485	Arg	Glu	Glu	Val	Ile 490	Asp	Phe
	Ser	Lys	Pro 495	Phe	Met	Ser	Leu	Gly 500	Ile	Ser	Ile	Met	Ile 505	Lys	Lys	Pro
10	Gln	Lув 510	Ser	ГÀв	Pro	Gly	<b>Val</b> 515	Phe	Ser	Phe	Leu	Asp 520	Pro	Leu	Ala	Tyr
	Glu 525	Ile	Trp	Met	Сув	Ile 530	Val	Phe	Ala	Tyr	Ile 535	Gly	Val	Ser	Val	Val 540
15	Leu	Phe	Leu	Val	Ser 545	Arg	Phe	Ser	Pro	Tyr 550	Glu	Trp	His	Thr	Glu 555	Glu
-	Phe	Glu	Asp	Gly 560	Arg	Glu	Thr	Gln	Ser 565		Glu	Ser	Thr	Asn 570	Glu	Phe -
20	Gly	Ile	Phe 575	Asn	Ser	Leu	Trp	Phe 580	Ser	Leu	Gly	Ala	Phe 585	Met	Arg	Gln
	Gly	Сув 590	Хsр	Ile	Ser	Pro	Arg 595	Ser	Leu	Ser	Gly	Arg 600	Ile	Val	Gly	Gly
25	Val 605	Trp	Trp	Phe	Phe	Thr 610	Leu	Ile	Ile	Ile	Ser 615	Ser	Tyr	Thr	Ala	Asn 620
•	Leu	Ala	Ala	Phe	Leu 625	Thr	Val	Glu	Arg	Met 630	Val	Ser	Pro	Ile	Glu 635	Ser
30	Ala	Glu	Авр	Leu 640	Ser	Lys	Gln	Thr	Glu 645	Ile	Ala	Tyr	Gly	Thr 650	Leu	Авр
	Ser	Gly	Ser 655	Thr	Lys	Glu	Phe	Phe 660	Arg	Arg	Ser	Lys	Ile 665	Ala	Val	Phe
35	Asp	Lys 670	Met	Trp	Thr	Tyr	Met 675	Arg	Ser	Ala	Glu	Pro 680	Ser	Val	Phe	Val
	Arg 685	Thr	Thr	Ala	Glu	Gly 690	Val	Ala	Arg	Val	Arg 695	Lys	Ser	Lys	Gly	Lys 700
40	Tyr	Ala	Tyr	Leu	Leu 705	Glu	Ser	Thr	Met	Asn 710	Glu	Tyr	Ile	Glu	Gln 715	Arg
	Lys	Pro	Сув	Asp 720	Thr	Met	Lys	Val	Gly 725	Gly	Asn	Leu	Asp	Ser 730	Lys	Gly
45	Tyr	Gly	Ile 735	Ala	Thr	Pro	Lys	Gly 740	Ser	Ser	Leu	Gly	Thr 745	Pro	Val	Asn
	Leu	Ala 750	Val	Leu	Lys	Leu	.Ser 755	Glu	Gln	Gly	Val	Leu 760	Asp	Lys	Leu	Lys
50 . `	Asn 765	Lys	Trp	Trp	Tyr	Asp 770	ГÀй	Gly	Glu	Сув	Gly 775	Ala	Lys	Asp	Ser	Gly 780
	Ser	Lys	Glu	Lys	Thr 785	Ser	Ala	Leu	Ser	<b>Le</b> u 790		Asn	Val	Ala	Gly 795	Val
55	Phe	Tyr	Ile	Leu 800	Val	Gly	Gly	Leu	Gly 805	Leu	Ala	Met	Ĺeu	Val 810	Ala	Leu

5			815	-,-	-,-	-,-		820		. 010	, wra	гра	825		Lys	Val	
	Ala	Lys 830	Asn	Ala	G1n	Aen	11e 835	Asn	Pro	Sez	Ser	Ser 840	Gln	Asn	Ser	Glń	
10	Asn 845	Phe	Ala	Thr	Tyr	Lys 850	Glu	Gly	Tyr	Asn	Val 855	Tyr	Gly	Ile	Glu	Ser 860	
	Val	Lys	Ile					,									
	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO:5	:								
15		(i)	() () ()	QUEN A) L B) T C) S D) T	engt Ype: Tran	H: 2 nuc DEDN	761 leic BSS:	base aci dou	pai d	rs							
20		(ii)	) MOI	LECU	LE T	YPE:	CDN.	A ,									
÷		(ix)	(1	ATURI A) Ni B) Lo	AME/	KEY: ION:	sig 79.	_pep	tide								
25		(ix)	(2	ATURI A) Ni B) Lo	NE/								-				
30		(ix)	(2	ATURI A) Ni B) Lo	AME/	KEY: ION:	CDS	. 274	5							•	
		(xi)	SEÇ	QUENC	CE DI	escr:	IPTIO	ON: :	SEO :	ID N	0:5:			•			
	GAAT											AGC	rtcg:	TTT :	TAGG	CGTAGC	60
35						ATG	GGG	CAA	AGC	GTG	CTC	cee	aca	CTC	TTC Phe	777	111
40	TTA Leu	GTC Val -10	CTG Leu	GGG Gly	CTT Leu	TTG Leu	GGT Gly -5	CAT His	TCT Ser	CAC	GGA Gly	GGA Gly 1	TTC Phe	CCC Pro	AAC Asn	ACC Thr 5	159
	ATC Ile	AGC Ser	ATA Ile	GGT Gly	GGA Gly 10	CTT Leu	TTC Phe	ATG Het	AGA Arg	AAC Asn 15	ACA Thr	GTG Val	CAG Gln	GAG Glu	CAC His 20	AGC Ser	207
45	GCT Ala		AL Y	25	VIG	val	GIR	Leu	30	Asn	Thr	Asn	Gln	Asn 35	Thr	Thr	255
50	GAG Glu	гåв	40	Pne	nıs	Leu	ABN	45	His	Val	Asp	His	Leu 50	Asp	Ser	Ser	303
<b></b>	AAT Asn	AGT Ser 55	TTT Phe	TCC Ser	GTG Val	ACA Thr	AAT Aan 60	GCT Ala	TTC Phe	TGC Cyn	TCC Ser	CAG Gln 65	TTC Phe	TCG Ser	AGA Arg	GGG Gly	351
55	GTG Val 70	ıyr	GCC Ala	ATC Ile	TTT Phe	GGA Gly 75	TTC Phe	TAT Tyr	GAC Asp	Gln	ATG Met 80	Ser	ATG Met	AAC Asn	ACC Thr	CTG Leu	399

5	ACC Thr	TCC Ser	TTC Phe	TGT Cys	GGG Gly 90	GCC Ala	CTG Leu	CAC His	ACA Thr	TCC Ser 95	TTT Phe	GTT Val	ACG Thr	CCT Pro	AGC Ser 100	TTC Phe	4	447
	CCC Pro	ACT Thr	GAC Asp	GCA Ala 105	GAT Asp	GTG Val	CAG Gln	TTT Phe	GTC Val 110	ATC Ile	CAG Gln	ATG Met	CGC Arg	CCA Pro 115	caa	TTG Leu	. 4	195
10	AAG Lys	GGC Gly	GCT Ala 120	ATT Ile	CTG Leu	AGT Ser	CTT Leu	CTG Leu 125	GGT Gly	CAT His	TAC Tyr	AAG Lys	TGG Trp 130	GAG Glu	AAG Lyb	ITT Phe	5	343
15	GTG Val	TAC Tyr 135	CTC Leu	TAT	GAC Asp	ACA Thr	GAA Glu 140	CGA Arg	GGA Gly	TTT Phe	TCC	ATC Ile 145	CTC	CAA Gln	GCG Ala	ATT Ile	5	591
	150	GAA Glu	ALA	Ala	Val	Gln 155	Asn	Asn	Trp	Gln	Val 160	Thr	Ala	Arg	Ser	Val 165		539
20		Asn	116	rys	170	Val	Gln	Glu	Phe	Arg 175	Arg	Ile	Ile	Glu	Glu 180	Met	•	87
25	Asp	AGG Arg	vrd	185	GIU	Lys	Arg	Tyr	Leu 190	Ile	ysb	Сув	Glu	Val 195	Glu	Arg	7	735
	IIe	AAC Asn	200	116	Leu	Glu	Gln	Val 205	Val	Ile	Leu	Gly	Lys 210	His	Ser	Arg	7	83
30	GIĀ	TAT Tyr 215	H18	Tyr	Met	Leu	Ala 220	Asn	Leu	Gly	Phe	Thr 225	qaA	Ile	Leu	Leu	8	331
35	230	AGA Arg	AT	Met	H18	G1y 235	Gly	Ala	Aen	Ile	Thr 240	Gly	Phe	Gln	Ile	Val 245		379
•	ASN	AAT Asn	GIU	ABR	250	Het	Val	Gln	Gln	Phe 255	Ile	Gln	Arg	Trp	Val 260	Arg	9	27
40	Leu	GAT Asp	Glu	Arg 265	Glu	Phe	Pro	Glu	Ala 270	Lys	Asn	Ala	Pro	Leu 275	Lys	Tyr		75
	THE		280	Leu	Thr	H18	Asp	Ala 285	Ile	Leu	Val	Ile	Ala 290	Glu	Ala	Phe	10	23
<b>45</b>	Arg	TAC Tyr 295	Leu	Arg	Arg	Gln	Arg 300	Val	Asp	Val	Ser	Arg 305	Arg	Gly	Ser	Ala	10	71
50	310	GAC Asp	Сув	Leu	Ala	315	Pro	Ala	Val	Pro	1rp 320	Ser	Gln	GŢĀ	Ile	<b>Авр</b> 325		.19
	116	GAG Glu	Arg	ALE	330	Lys	Xet	Val	Gln	Val 335	Gln	Gly	Met	Thr	Gly 340	Asn	11	.67
55	ATT	CAA Gln	TTT Phe	GAC Asp 345	ACT Thr	TAT Tyr	GGA Gly	CGT Arg	AGG Arg 350	ACA Thr	AAT Asn	TAT Tyr	ACC Thr	ATC Ile 355	GAT Asp	GTG Val	12	15

5									CGA Arg								;	1263
40									GAT Asp									1311
10									GTA Val								•	1359
15									CAT His									1407
									CTA Leu 430									1455 -
20									ATC Ile									1503
25	Ala	Arg 455	Asp	Pro	Glu	Thr	Lys 460	Ile	TGG Trp	Asn	Gly	Met 465	Val	Gly	Glu	Leu		1551
									GTT Val									1599
30	GTC Val								TCA Ser							Gly		1647
35									Gln 510									1695
									GAA Glu	_				Ile				1743
40			Ile					Val	CTT Leu									1791
45		Tyr										Glu				Pro-		1839
45						Pro					Gly					CTT Leu		1887
50					Gly					Gln					Ser	CCA Pro		1935
				Ser					Gly					Phe		ACC Thr		1983
55			Ile					Thr					Ala			ACT Thr		2031

5									GAG Glu								2079
									CTG Leu								2127
10									GTG Val 670								2175
15									TTT Phe								2223
									GGA Gly								2271
20									CAG Gln								2319_
25									AAA Lys								2367
									GTT Val 750								2415
30									TTG Leu					Trp			2463
35			Glu						GGT Gly							AGC Ser	2511
35		Leu							GGC							GGA Gly 805	2559
40						Het			GCT Ala		Ile						2607
					Ser					Leu					Gln	AAC Asn	2655
<b>4</b> 5				Ala					Thr					Thr		AGA Arg	2703
50	GAA Glu	GGC Gly 855	Tyr	AAC Asn	GTG Val	TAT	GGA Gly 860	Thr	GAG Glu	AGT Ser	GTT Val	AAG Lys 865	Ile	TAG	GGAT	ccc	2752
	TTG	GAAT	TC														2761

(2)	INFORMATION	FOR	SEQ	ID	NO:	6	:
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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 888 amino acids
  - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Gly Gln Ser Val Leu Arg Ala Val Phe Phe Leu Val Leu Gly Leu -22 -20 -15 -10

Leu Gly His Ser His Gly Gly Phe Pro Asn Thr Ile Ser Ile Gly Gly -5 5 10

Leu Phe Met Arg Asn Thr Val Glu His Ser Ala Phe Arg Phe Ala 15 20 25

Val Gln Leu Tyr Asn Thr Asn Gln Asn Thr Thr Glu Lys Pro Phe His 30 35 40

Leu Asn Tyr His Val Asp His Leu Asp Ser Ser Asn Ser Phe Ser Val 45 50 55

Thr Asn Ala Phe Cys Ser Gln Phe Ser Arg Gly Val Tyr Ala Ile Phe 60 65 70

Gly Phe Tyr Asp Gln Met Ser Met Asn Thr Leu Thr Ser Phe Cys Gly 75 80 85 90

Ala Leu His Thr Ser Phe Val Thr Pro Ser Phe Pro Thr Asp Ala Asp 95 100 105

Val Gln Phe Val Ile Gln Met Arg Pro Ala Leu Lys Gly Ala Ile Leu 110 115 120

Ser Leu Leu Gly His Tyr Lys Trp Glu Lys Phe Val Tyr Leu Tyr Asp 125 130 135

Thr Glu Arg Gly Phe Ser Ile Leu Gln Ala Ile Met Glu Ala Ala Val 140 145 150

Gln Asn Asn Trp Gln Val Thr Ala Arg Ser Val Gly Asn Ile Lys Asp 155 160 165 170

Val Gln Glu Phe Arg Arg Ile Ile Glu Glu Met Asp Arg Arg Gln Glu 175 180 185

Lys Arg Tyr Leu Ile Asp Cys Glu Val Glu Arg Ile Asn Thr Ile Leu 190 195 200

Glu Gln Val Val Ile Leu Gly Lys His Ser Arg Gly Tyr His Tyr Met 205 210 215

Leu Ala Asn Leu Gly Phe Thr Asp Ile Leu Leu Glu Arg Val Met His 220 225 230

Gly Gly Ala Asn Ile Thr Gly Phe Gln Ile Val Asn Asn Glu Asn Pro 235 240 250

Met Val Gln Gln Phe Ile Gln Arg Trp Val Arg Leu Asp Glu Arg Glu 265 265

Phe Pro Glu Ala Lys Asn Ala Pro Leu Lys Tyr Thr Ser Ala Leu Thr 270 275 280

	His	Asp	Ala 285	Ile	Leu	Vál	Ile	Ala 290	Glu	Ala	Phe	Arg	Tyr 295	Leu	Arg	Arg
5	Gln	Arg 300	Val	Asp	Val	Ser	Arg 305	Arg	Gly	Ser	Ala	Gly 310	Asp	Сув	Leu	Ala
	Asn 315	Pro	Ala	Val	Pro	Trp 320	Ser	Gln	Gly	Ile	Asp 325	Ile	Glu	Arg	Ala	Leu 330
10	Lys	Met	Val	Gln	Val 335	Gln	Gly	Met	Thr	Gly 340	Asn	Ile	Gln	Phe	Asp 345	Thr
				Arg 350					355					360	_	
15			365	Arg				370					375			
		380		Asp			385					390				
20	395			Val		400					405	•	-			410
				His	415					420					425	
25				Leu 430					435					440	_	-
			445	Ile				450					455			
30		460		Trp			465					470	-	_	_	
	475			Val		480					485					490
35				Ser	495					500					505	
				Gln 510					515					520	=	
40			525	Glu				530					535		-	
	•	540		Leu			545					550			_	
45	555			Asn		560					565					570
				Glu	575					580					585	
50				Gln 590					595					600		
			605	Gly				610					615			
55	Ser	Tyr 620	Thr	Ala	Asn	Leu	Ala 625	Ala	Phe	Leu	Thr	Val 630	Glu	Arg	Met	Val

	Ser 635	Pro	Ile	Glu	Ser	Ala 640	Glu	Asp	Leu	Ala	Lys 645	Gln	Thr	Glu	Ile	Ala 650
5	Tyr	Gly	Thr	Leu	<b>Asp</b> 655	Ser	Gly	Ser	Thr	Lys 660	Glu	Phe	Phe	Arg	Arg 665	Ser
	Lys	Ile	Ala	Val 670	Tyr	Glu	Lys	Met	Trp 675	Ser	Tyr	Met	Lys	Ser 680	Ala	Glu
10	Pro	Ser	Val 685	Phe	Thr	Lys	Thr	Thr 690	Ala	Asp	Gly	Val	Ala 695	Arg	Val	Arg
	Lys	Ser 700	Lys	Gly	Lys	Phe	Ala 705	Phe	Leu	Leu	Glu	Ser 710		Met	Asn	Glu
15	Tyr 715	Ile	Glu	Gln	Arg	Lys 720	Pro	Сув	Asp	Thr	Met 725	Lys	Val	Cly	Gly	Asn 730
	Leu	Asp	Ser	Lys	Gly 735	Tyr	Gly	Val	Ala	Thr 740	Pro	Lys	Gly	Ser	Ala 745	Leu
20	Gly	Asn	Ala	Val 750	Asn	Leu	Ala	Val	Leu 755	Lys	Leu	Asn	Glu	Gln 760	Gly	Leu
	Leu	qBA	Lys 765	Leu	Lys	Asn	Lys	Trp 770	Trp	Tyr	Asp	Lys	Gly 775	Glu	Сув	Gly
25	Ser	Gly 780	Gly	Gly	Asp	Ser	Lys 785	Asp	Lys	Thr	Ser	Ala 790	Leu	Ser	Leu	Ser
	Asn 795	Val	Ala	Gly	Val	Phe 800		Ile	Leu	Val	Gly 805	Gly	Leu	Gly	Leu	Ala 810
30	Met	Met	Val	Ala	Leu 815	Ile	Glu	Phe	Сув	Tyr 820	Lys	Ser	Arg	Ala	Glu 825	
	Lys	Arg	Met	Lys 830	Leu	Thr	Lys	Asn	Thr 835		Asn	Phe	Lys	Pro 840		Pro
35	Ala	Thr	Asn 845	Thr	Gln	Asn	Tyr	Ala 850		Tyr	Arg	Glu	Gly 855		Asn	Val
	Tyr	Gly 860		Glu	Ser	Val	Lys 865	Ile								
40	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO: 7	•							
		(1	<b>'</b> (	Ã) L	ENGT	H: 3	CTER	base	pai	.rs		•				
45			į	c) s	TRAN	DEDN	leic ESS: lin	dou								
		(ii	.) <b>K</b> O	LECU	LE I	YPE:	cDN	A		·						
50		(ix	· (	ATUR A) N B) L	AME/	KEY:	: sig	_per	etide 							
55		ĸi)	(		IAME/		mat			•						

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(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 79..2745

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

		(x1)	SEQ	UENC	E DE	SCRI	PTIU	N: 2	EQ I	טא ט.	): /:							
10	GAAT	TCCT	GA C	GACT	CCTG	A GI	TGCG	CCCA	TGC	TCTI	GTC	AGCT	TCGT	TT T	AGGC	GTAGC		60
,,	ATGG	CCAG	GC A	GAAĞ								CGG Arg						111
15												GGA Gly 1						159
												GTG Val				AGC Ser		207
20												AAC Asn					٠	255
25												CAC His						303
												CAG Gln 65						351
30	GTG Val 70	TAT Tyr	GCC Ala	ATC Ile	TTT Phe	GGA Gly 75	TTC Phe	TAT Tyr	GAC Asp	CAG Gln	ATG Met 80	TCA Ser	ATG Met	AAC Asn	ACC Thr	CTG Leu 85		399
35												GTT Val						447
~												ATG Met						495
40	AAG Lys	GGC Gly	GCT Ala 120	ATT Ile	CTG Leu	AGT Ser	CTT Leu	CTG Leu 125	GGT Gly	CAT His	TAC	AAG Lys	TGG Trp 130	Glu	AAG Lys	TTT Phe		543
			Leu					Arg				ATC Ile 145	Leu			ATT	•	591
45		Glu					Asn					ACA Thr						639
50						Val					Arg					ATG Het		687
					Glu					Ile					Glu	AGG Arg	•	735

55

5		AAC Asn															783
10		TAT Tyr 215															831
-		AGA Arg															879
15		AAT Asn														λGG Arg	927
		GAT Asp															975
20		TCT Ser															1023
25		TAC Tyr 295															1071
		GAC Asp															1119
30		GAG Glu														Asn	1167
35		CAA Gln															1215
				Lys					Arg					Trp		GAG Glu	1263
40 ·			Arg										Ser			AGT Ser	1311
		Ser					Thr					Thr				TCA Ser 405	1359
45	CCA Pro	TAT	GTA Val	ATG Met	TAC Tyr 410	Lys	AAG Lys	AAC	CAT	GAG Glu 415	Gln	CTG	GAA Glu	GGA Gly	AAT ABD 420	GAA Glu	1407
50	CGA Arg	TAT Tyr	GAA Glu	GGC Gly 425	Tyr	TGT Cys	GTA Val	GAC Asp	Leu 430	Ala	TAT	GAA Glu	ATA Ile	GCC Ala 435	Lys	CAT His	1455
	GTA Val	AGG Arg	Ile 440	Lys	TAC	Lys	TTG Leu	TCC Ser 445	Ile	GTT Val	GGT	Asp Asp	GGG Gly 450	Lys	TAT Tyr	GGT Gly	1503
55			Asp					Ile					. Val			CTT	

5	GTC Val 470	TAT Tyr	G1y	AGA Arg	GCT Ala	GAT Asp 475	ATA Ile	GCT Ala	GTT Val	Ala	CCA Pro 480	CTC Leu	ACT Thr	ATA Ile	ACA Thr	TTG Leu 485	1599
	GTC Val	CGT Arg	GAA Glu	GAA Glu	GTC Val 490	ATA Ile	GAT Asp	TTT Phe	TCA Ser	AAG Lys 495	CCA Pro	TTA Leu	ATG Met	AGC Ser	CTG Leu 500	GGC Gly	1647
10	ATC Ile	TCC Ser	ATC Ile	ATG Met 505	ATA Ile	AAG Lys	AAG Lys	CCT Pro	CAG Gln 510	AAA Lys	TCA Ser	AAA Lys	CCA Pro	GGC Gly 515	GTA Val	TTC Phe	1695
15	TCA Ser	TTT Phe	CTG Leu 520	GAT Asp	CCC Pro	CTG Leu	GCT Ala	TAT Tyr 525	GAA Glu	ATC Ile	TGG Trp	ATG Ket	TGC Cys 530	ATT	GTC Val	TTT Phe	1743
	GCT Ala	TAC Tyr 535	ATT Ile	GGA Gly	GTC Val	AGC Ser	GTA Val 540	GTT Val	CTT Leu	TTC Phe	CTA Leu	GTC Val 545	AGC Ser	AGG Arg	TTC Phe	AGT Ser	1791
20	CCT Pro 550	TAT Tyr	GAA Glu	TGG Trp	CAC His	TTG Leu 555	GAA Glu	GAC Asp	AAC Asn	AAT Asn	GAA Glu 560	Glu	CCT Pro	CGT	GAC	CCA Pro 565	1839
25	CAA Gln	AGT Ser	CCT Pro	CCT Pro	GAT Asp 570	CCT Pro	CCA Pro	AAT Asn	GAA Glu	TTT Phe 575	GGA Gly	ATA Ile	TTT Phe	AAC Asn	AGT Ser 580	Leu	1887
	TGG Trp	TTT Phe	TCC Ser	TTG Leu 585	GGT	GCC Ala	TTT Phe	ATG Met	CAG Gln 590	CAA Gln	GGA Gly	TGT Cys	GAT Asp	ATT Ile 595	Ser	Pro	1935
30	AGA Arg	TCA Ser	CTC Leu 600	Ser	GGG	CGC Arg	ATT	GTT Val 605	Gly	GGG	GTT Val	TGG	TGG Trp 610	Phe	TTC Phe	ACC Thr	1983
35	CTG Leu	ATC Ile 615	Ile	ATT	TCT Ser	TCC Ser	TAT Tyr 620	Thr	GCC	AAT Asn	CTC	GCT Ala 625	. Ala	TTC Phe	Lev	ACT	2031
33	GTG Val 630	Glu	AGG Arg	ATG Met	GTT Val	TCT Ser 635	Pro	ATA Ile	GAG Glu	AGT Ser	GCT Ala 640	Glu	GAC Asp	TTA Lev	GC1	C AAA Lys 645	2079
40	CAG Gln	ACT Thi	GAA Glu	ATT	GCA Ala 650	Tyr	GCG	ACC Thr	CTG Leu	GAC Asp 655	Ser	GGT Gly	TCA Ser	AC:	Lyi 660	A GAA B Glu	2127
	TTT	TTC Phe	AGA Arg	AGA Arg 665	Ser	AAA Lys	ATT	GCT	GTG Val 670	Tyr	GAC Glu	AAA Lys	A ATC	TG(	Se	TAC Tyr	2175
45	ATC Met	Ly	A TCA Ser 680	Ala	GAG Glu	CCA Pro	TCI Ser	GTG Val 685	Phe	ACC Thi	Lyi	A AC	A ACI	r Al	A GA	c GGA p Gly	2223
50	GTC Val	GC( Al-	a Arg	GTC Val	CGA	A AAG J Lys	Sex 700	: Lys	GG; Gl <sub>j</sub>	A AAC 7 Lyi	TTO Pho	C GCG B Ala 70	a Pho	C CT	G CT	G GAG u Glu	2271
	TCI Sei 710	Th	C ATO	3 AA: : AB:	r GAG	TAC 1 Tyl 715	: Ile	GAG	CAC 1 Gl	AGI Ar	A AAI g Ly: 72	8 Pr	A TG	T GA B AB	T AC p Th	G ATG r Met 725	2319
55	AAJ Lyi	A GT s Va	r GG? l Gl;	r GGI	A AA: 7 As: 73	n Lei	GA:	r TC	C AAI	A GG 6 G1 73	y Ty	T GG r Gl	T GT Y Va	G GC l Al	A AC a Th 74	C CCT r Pro 0	2367

5	AAA Lys	GGC	TCA Ser	GCA Ala 745	TTA Leu	GGA Gly	ACG Thr	CCT Pro	GTA Val 750	AAC Asn	CTT Leu	GCA Ala	GTA Val	TTG Leu 755	AAA Lys	CTC Leu	2415
	AGT Ser	GAA Glu	CAA Gln 760	GGC Gly	ATC Ile	TTA Leu	GAC Asp	AAG Lys 765	CTG Leu	AAA Lys	AAC Asn	AAA Lys	TGG Trp 770	TGG Trp	TAC Tyr	GAT Asp	2463
10	AAG Lys	GGG Gly 775	GAA Glu	TGT Cys	GGA Gly	GCC Ala	AAG Lys 780	GAC Asp	TCC Ser	GGG Gly	AGT Ser	AAG Lys 785	GAC Asp	AAG Lys	ACC Thr	AGC Ser	2511
15	GCT Ala 790	CTG Leu	AGC Ser	CTG Leu	AGC Ser	AAT Asn 795	GTG Val	GCA Ala	GGC Gly	GTT Val	TTC Phe 800	TAT Tyr	ATA Ile	CTT Leu	GTC Val	GGA Gly 805	2559
	GGT Gly	CTG Leu	GGG Gly	CTG Leu	GCC Ala 810	ATG Met	ATG Ket	GTG Val	GCT Ala	TTG Leu 815	ATA Ile	GAA Glu	TTC Phe	TGT Cys	TAC Tyr 820	AAA Lys	2607
20	TCA Ser	CGG Arg	GCA Ala	GAG Glu 825	TCC Ser	AAA Lys	CGC Arg	ATG Met	AAA Lys 830	CTC Leu	ACA Thr	AAG Lys	AAC Asn	ACC Thr 835	CAA Gln	AAC Asn	2655
25	TTT Phe	AAG Lye	CCT Pro 840	GCT Ala	CCT Pro	GCC Ala	ACC Thr	AAC Asn 845	ACT Thr	CAG Gln	AAT Asn	TAT Tyr	GCT Ala 850	ACA Thr	TAC Tyr	AGA Arg	2703
	GAA Glu	GGC Gly 855	TAC Tyr	AAC Asn	GTG Val	TAT Tyr	GGA Gly 860	ACA Thr	GAG Glu	AGT Ser	GTT Val	AAG Lys 865	ATC Ile	TAG	GGAT(	ccc	2752
30	TTC	CCACT	rgg 2	AGGC	ATGT	A TO	AGAC	GAAI	A TC	ACCG	AAA	CGT	GCT	GCT '	TCAA	GGATCC	2812
	TGA	CCAC	AT :	PTCAC	CTCTC	CC T	rggto	STCG	GC	ATGA	CACG	AAT	ATTG	CTG :	ATGG	TGCAAT	2872
	GAC	CTTT	CAA !	[agġi	LAAA	AC TO	ATT	CTTT:	r TT:	rcct:	CAG	TGC	CTTA	rgg :	AACA	CTCTGA	2932
35	GAC	rcgco	SAC 2	AATGO	CAAAC	CC A	CAT	rgaai	A TC	rttt:	rgct	TTG	CTTG:	AAA .	AAAA	ATAATT	2992
<b>3</b> 5	AAA	LAATA	AAA (	CCAAC	LAAAC	AA TO	GAC	ATGC:	A TC	AAAC	CCTT	GAT	STAT	TAA '	TATT	TATTAT	3052
	AGT	rttc/	ATT I	AGGAI	ATTC			-									3070
40	(2)	INFO	ORMA!	rion	FOR	SEO	ID 1	NO: 8									
••	, ,			SEQUI (A) (B)	ENCE	CHAI NGTH PE:	RACTI 880	ERIS	TICS ino a	: Acid	В						
45		(	Li) 1	MOLE	CULE	TYP	E: p	rote.	in			•					
		(2	ci) :	SEQUI	ENCE	DES	CRIP	LION	: SE	Q ID	NO:	8:					
50	Met -22	Gly	Gln -20	Ser	Val	Leu	Arg	λla -15	Val	Phe	Phe	Leu	Val -10	Leu	Gly	Leu	
	Leu	Gly -5	His	Ser	His	Gly	Gly 1	Phe	Pro	Asn	Thr 5	Ile	Ser	Ile	Gly	Gly 10	
	Leu	Phe	Met	Arg	Asn 15	Thr	Val	Gln	Glu	His 20		Ala	Phe	Arg	Phe 25		

Val Gln Leu Tyr Asn Thr Asn Gln Asn Thr Thr Glu Lys Pro Phe His 30 40

	Leu	Asn	Tyr 45	His	Val	Asp	His	Leu 50	qaA	Ser	Ser	Asn	Ser 55	Phe	Ser	Val
5	Thr	Asn 60	Ala	Phe	Сув	Ser	Gln 65	Phe	Ser	Arg	Gly	<b>Val</b> 70	Tyr	Ala	Ile	Phe
	Gly 75	Phe	Tyr	Asp	Gln	Met 80	Ser	Met	Asn	Thr	Leu 85	Thr	Ser	Phe	Сув	Gly 90
10	Ala	Leu	His	Thr	Ser 95	Phe	Val	Thr	Pro	Ser 100	Phe	Pro	Thr	qaA	Ala 105	Asp
	Val	Gln	Phe	Val 110	Ile	Gln	Met	Arg	Pro 115	Ala	Leu	, Lys	Gly	Ala 120	Ile	Leu
- 15	Ser	Leu	Leu 125	Gly	His	Tyr	Lys	Trp 130	Glu	Lys	Phe	Val	Tyr 135	Leu	Tyr	Asp
	Thr	Glu 140	Arg	Gly	Phe	Ser	Ile 145	Leu	Gln	Ala	Ile	Met 150	Glu	Ala	Ala	Val·
20	. Gln 155	Asn	Asn	Trp	Gln	Val 160	Thr	Ala	Arg	Ser	Val 165	Gly	Asn	Ile	Lys	Asp 170
	Val	Gln	Glu	Phe	Arg 175	Arg	Ile	Ile	Glu	Glu 180	Met	Asp	Arg	Arg	Gln 185	Glu
25	Lys	Arg	Tyr	Leu 190	Ile	Asp	Сув	Glu	Val 195	Glu	Arg	Ile	Asn	Thr. 200	Ile	Leu
	Glu		Val 205	Val.	Ile	Leu	Gly	Lys 210	His	Ser	Arg	Gly	Tyr 215	His	Tyr	Met
30	Leu	Ala 220	Asn	Leu	Gly	Phe	Thr 225	Авр	Ile	Leu	Leu	Glu 230	Arg	Val	Met	His
	Gly 235	Gly	Ala	Asn	Ile	Thr 240	Gly	Phe	Gln	Ile	Val 245	Asn	Asn	Glu	Asn	Pro 250
35	Met	Val	Gln	Gln	Phe 255	Ile	Gln	Arg	Trp	Val 260	Arg	Leu	Asp	Glu	Arg 265	Glu
	Phe	Pró	Glu	Ala 270	Lys	Asn	Ala	Pro	Leu 275	Lys	Tyr	Thr	Ser	Ala 280	Leu	Thr
40	His	Asp	Ala 285		Leu	Val	Ile	Ala 290	Glu	Ala	Phe	Arg	Tyr 295		Arg	Arg
	Gln	Arg 300		Asp	Val	Ser	Arg 305	Arg	Gly	Ser	Ala	Gly 310	Asp	Сув	Leu	Ala
45	Asn 315		Ala	Val	Pro	Trp 320		Gln	Gly	Ile	Авр 325		Glu	Arg	λla	<b>Leu</b> 330
	Lys	Met	Val	Gln	Val 335		Gly	Met	Thr	Gly 340		Ile	Gln	Phe	Авр 345	Thr
50	Tyr	Gly	Arg	Arg 350		Asn	Tyr	Thr	11e 355		Val	Tyr	Glu	. Met 360		Val
	Ser	Gly	Ser 365		Lys	Ala	Gly	Tyr 370		Asn	Glu	Tyr	Glu 375		Phe	val
55	Pro	Phe 380		Asp	Gln	Gln	11e 385		Asn	Asp	Ser	Ala 390		Ser	Glu	Asn

	Arg 395	Thr	Ile	Val '		Th <i>r</i> 400	Thr	Ile	Leu	Glu	Ser 405	Pro	Tyr	Val	Met	Tyr 410
5	Lys	Lys	Asn		Glu 415	Gln	Leu	Glu	Gly	Asn 420	Glu	Arg	Tyr	Glu	Gly 425	Tyr
	Сув	Val	Asp	Leu 430	Ala	Tyr	Glu	Ile	Ala 435	Lys	His	Val	Arg	Ile 440	Lys	Tyr
10	Lys	Leu	Ser 445	Ile	Val	Gly	Asp	Gly 450	Lys	Tyr	Gly	Ala	Arg 455	Asp	Pro	Glu
	Thr	Lys 460	Ile	Trp	naA	Gly	Met 465	Val	Gly	Glu	Leu	Val 470	Tyr	Gly	Arg	Ala
15	475			Val		480					485					490
				Ser	495		·			500					505	
20	_			Gln 510					515					520		
			525	Glu				530			·		535			
25		540		Leu			545			•		550				
	555			Asn		560					565					570
30				Glu	575					580	1				585	
				590					595	i				600		Gly
35	_		605		٠			610	)	•			615			Ser
		620					625	i				630	,			. Val
40	635	· ·				640	)				645	•				650
					655					660	)				66	
45	Lys	ı Ile	a Ala	670		Glu	ı Lys	Met	67!	p Sei 5	r Tyi	r Met	L Ly	68	r Ala	a Glu
	Pro	Ser	68!		Thr	Lyı	B . Thi	690		a As	p Gly	y Va.	1 Ala 69	a Ar	g Va	l Arg
50	-	700	)				70	5				71	0			n Glu
	71	5				72	0				72	5				730
55	Le	u As	p Se	r Ly	8 Gly		r Gl	y Va	l Al	a Th 74	r Pr O	o Ly	s Gl	y Se	r Al 74	a Leu 5

	Gly	Thr	Pro	Val 750	Asn	Leu	Ala	Val	Leu 755	Lys	Leu	Ser	Glu	Gln 760	Gly	Ile	
5	Leu	Asp	Lys 765	Leu	Lys	Asn	Lys	Trp 770	Trp	Tyr	yab	Lys	Gly 775	Glu	Сув	Gly	
	Ala	Lys 780	Asp	Ser	Gly	Ser	Lys 785	двр	Lys	Thr	Ser	Ala 790	Leu	Ser	Leu	Ser	
10	Asn 795	Val	Ala	Gly	Val	Phe 800	Tyr	Ile	Leu	Val	Gly 805	Gly	Leu	Gly	Leu	Ala 810	
	Met	Met	Val	Ala	Leu 815	Ile	Glu	Phe	Сув	Tyr 820	Lув	Ser	Arg	Ala	Glu 825	Ser	
15	Lys	Arg	Met	Lys 830	Leu	Thr	Lys	Asn	Thr 835	Gln	Asn	Phe	Lys	Pro 840	Ala	Pro	
	Ala	Thr	Asn 845	Thr	Gln	Asn	Tyr	Ala 850	Thr	Tyr	Arg	Glu	Gly 855	Tyr	Asn	Val ·	
20	Tyr	Gly 860	Thr	Glu	Ser	Val	Lys 865	Ile									
<b>25</b>	(2)		SE(	QUEN A) L	FOR CE CI ENGTI YPE:	HARA H: 4	CTER 6 am	ISTI	cs:	8							
30		(ii)	·		opol				·								
35		•	•	_	CE D a Le				_				a.Va	ıl Le	u Ly	s Leu 15	Asn
		Gl	u Gl	n Gl	y Le 20		u As	p Ly	s Le	u Ly 25		n Ly	s Tr	p Tr	ъ Ту 30	r Asp	Lys
40		G1	y Gl	u Cy 35	s Gl	y Se	r Gl	y Gl	y Gl 40		p Se	r Ly	s As	p Ly 45		r	
	(2)				FOR												
45		(1	( (	Ā) L B) T	CE C ENGT YPE: OPOL	H: 4	no a	ino		ls							
		(11	) нс	LECU	ILE T	YPE:	per	tide	•								
50		(×i	.j SE	:QUEN	ice i	ESCF	RIPT	ON:	SEQ	ID N	10:10	):					
		G1 1	y Se	er Al	la Le	su G1 5	Ly Th	nr Pr	:o . Va	al As	n Le		La V	al L	au L	ys Leu 15	Ser
55		G1	.u Gl	in Gl	Ly II 20		eu As	sp Ly	/B Le	eu Ly 25	/8 As	n L	/8 T	rp T	rp T	yr Asg O	Lys

5	Giy Glu Cys Giy Ala Lys Asp Ser Gly Ser Lys Asp Lys Thr 35 40 45	
	(2) INFORMATION FOR SEQ ID NO:11:	
10	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 13 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: Other nucleic acid; (A) DESCRIPTION: Synthetic DNA oligonucleotide	
15	•	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
	AGCTTGCGGC CGC	13
20	(2) INFORMATION FOR SEQ ID NO:12:	
25	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 9 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
25	<ul><li>(ii) MOLECULE TYPE: Other nucleic acid;</li><li>(A) DESCRIPTION: Synthetic DNA oligonucleotide</li></ul>	
	(iv) ANTI-SENSE: YES	
30		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
	GCGGCCGCA	9
35	(2) INFORMATION FOR SEQ ID NO:13:	
	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 40 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
40	(ii) MOLECULE TYPE: Other nucleic acid; (A) DESCRIPTION: Synthetic DNA oligonucleotide	
45	(x1) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
	ACACTCAGAA TTACGCTACA TACAGAGAAG GCTACAACGT	40
•	(2) INFORMATION FOR SEQ ID NO:14:	
50	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 40 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
55	<ul><li>(ii) MOLECULE TYPE: Other nucleic acid;</li><li>(A) DESCRIPTION: Synthetic DNA oligonucleotide</li></ul>	

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
5	CCĄGATCGAT ATTGŤGAACA TCAGCGACAC GTTTGAGATG	40
	(2) INFORMATION FOR SEQ ID NO:15:	
	(i) SEQUENCE CHARACTERISTICS:	

(A) LENGTH: 32 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Other nucleic acid;
(A) DESCRIPTION: Synthetic DNA oligonucleotide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GTGAATGTGG AGCCAAGGAC TCGGGAAGTA AG

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### Claims

- An isolated polynucleotide comprising a region which encodes an AMPA-binding human GluR receptor selected from the group consisting of human GluR1B, GluR2B, GluR3A and GluR3B receptors, and AMPAbinding fragments thereof.
- An isolated polynucleotide according to claim 1, which encodes said GluR1B receptor, said GluR2B receptor, said GluR3A receptor or said GluR3B receptor.
  - 3. An isolated polynucleotide comprising a region which encodes an AMPA-binding variant of a GluR receptor selected from the group consisting of human GluR1B, GluR2B, GluR3A and GluR3B receptors, wherein said variant has the binding profile of said receptor and varies from said receptor by conservative amino acid substitution.
  - 4. An isolated polynucleotide according to any one of claims 1 to 3, which consists of DNA.
  - A recombinant DNA construct having incorporated therein a polynucleotide as defined in any one of claims 1 to 4.
  - A recombinant DNA construct according to claim 5, wherein the polynucleotide incorporated therein is linked operably with DNA enabling expression and secretion of said receptor in a cellular host.
- A recombinant DNA construct according to claim 5, which is plasmid pBS/humGluR3A (ATCC 75218), plasmid pBS/humGluR3B (ATCC 75219); plasmid pBS/humGluR1B (ATCC 75246) or plasmid pBS/humGluR2B (ATCC 75217).
  - A cellular host having incorporated therein a heterologous polynucleotide as defined in any one of claims 1 to 4.
  - 9. A cellular host according to claim 8, which is a mammalian cell.
  - 10. An AMPA-binding membrane preparation derived from a cellular host as defined in claim 8 or claim 9.
- 11. A process for obtaining a substantially homogeneous source of human GluR receptor, which comprises the step of culturing a cellular host as defined in claim 8 or claim 9, and then recovering the cells so cultured.
  - 12. A process for obtaining a substantially homogeneous source of human GluR receptor according to claim

11 comprising the subsequent step of obtaining a membrane preparation from the cultured cells.

13. A method of assaying a substance for binding to a human EAA receptor, which comprises the steps of incubating the substance under apprópriate conditions with a cellular host as defined in claim 8 or claim 9, or with an AMPA-binding membrane preparation derived therefrom, and determining the extent of binding between the human GluR receptor and the substance.

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- 14. An isolated human GluR receptor selected from the group consisting of GluR1B, GluR2B, GluR3A and GluR3B receptors, and AMPA-binding fragments thereof, in a form essentially free from other proteins of human origin.
  - An AMPA-binding fragment of a human GluR receptor selected from the group consisting of GluR1B, GluR2B, GluR3A and GluR3A receptors.
- 16. An antibody which binds a human GluR receptor selected from the group consisting of GluR1B, GluR2B, GluR3A and GluR3B receptors.
  - 17. An immunogenic fragment of a human GluR receptor selected from the group consisting of GluR1B, GluR2B, GluR3A and GluR3B receptors.
  - 18. An oligonucleotide which comprises at least about 17 nucleic acids and which hybridizes selectively with a polynucleotide defined in any one of claims 1 to 4.

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61	TAIGCAGCACATITITGCCTICTTCTGCACCGGTTTCCTAGGCGCGGTAGTAGGTGCCAA	AGC	ACA:	PTT	PTGC	CTT	CTT	CTG	CAC	000	TTI	CCL	AGG +	000	GGT	AGT	AGG	TGC	CAA	
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† †	ATAACACTTGTAGTCGCTGTGCAAACTCTACTGGATATCTAAGACAAGGGTCAAGAGGTT	ACTI	rgta	GTC	GCT	GTC	CAA	ACT	CTA	CTG	PAT	TC	GAA	SAC	NAGC	GTC	AA.	346	GTT	300
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TITGITITAAGITCCTCTCACCGGGTTACACTGTCCAAAGGTCGACCACTTGATGTGTCT  N K F K E S G A N V T G F Q L V N Y T D  CACTATTCCGGCCAAGATCATGCAGCAGTGGAAGAATAGTGATGCTCGAGACCACACG	K N G I G Y H Y I L A N L G F	CTTCTTACCGTAGCCGATGGTGATGTAAGAACGTTTAGACCCGAAGTACCTGTAACTGAA	ATGGCATCGGCTACCACTAC	臼	CCTGACACTTAGTCTTGCG	+	STGGACTGTGAATCAGAACGC		E E G Y "R M L	CTCCTCCTATGGCCTACGAGE

## F1G. 10

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1701	ACCCCTAACAGACCGATTGGGTCGACAAGGGACCCCGGTTCCCTAGCTGTAGGTCTCTCG	1080
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6	TCTGCAGCAGGTGCGATTTGAAGGTTTAACAGGAAACGTGCAGTTTAATGAGAAAGGACG	7
1001	AGACGICGICCACGCIAAACTICCAAATIGICCITIGCACGICAAATIACICITICCIGC	1140
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141	GGCCTGGTTGATGTGCGAGGTGCACTATACTTTGTACTGCCGTAGGCTTTCTAACC	7
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### F/G. 1E

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### F16. 1F

+++ 1560 CGACCAGATACCTTCTCGTCTACACCGACACCGAGGGAATTGATAGTGAAACCAGGCCCT	LVYGRADVAVAPLTITLVRE-	AGAAGTTATAGATTTCTCCAAACCATTTATGAGTTTGGGGATCTCCATCATGATTAAAA	TCTTCAATATCTAAAGAGGTTTGGTAAATACTCAAACCCCTAGAGGTAGTACTAATTTTT	VIDFSKPFMSLGISIMIKK-		TGGTGTCTTTAGGTTCGGCCCACAGAAGAGGAAGGAACTAGGAAACCGAATACTCTAAAC	P Q K S K P G V F S F L D P L A Y E I W -	GATGTGCATTGTTTTTGCCTACATTGGAGTGAGTGTTGTCCTCTTCCTGGTCAGCCGCTT		MCIVFAYIGVSVULFLVSRF-	CAGTCCCTATGAATGGCACAGTGAAGAGTTTGAGGAAGGA	1/41	1 C W F F C C C W E E E W H 3 E A W
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### F/G. 16

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1801+ GGTCAGGTTACT	S S S	GCAAGGATGTGA( 1861	Q G. C D	GTTCTTCACCTT/ 1921+ CAAGAAGTGGAAT	म स	GGAGAGGATGGTC 1981		E R M V	CTACGGGACGCTC	_	Y G T L

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1076	CCATGACTTCCCCAAGTCCATGCAATCGATTCCTTGCATGAGCCACAGTTCAGGGATGCC	0000
7077	GGTACTGAAGGGGTTCAGGTACGTTAGCTAAGGAACGTACTCGGTGTCAAGTCCCTACGG	09/7
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	CTTGGGAGCCACGGGATTGTAACTGGAGCAGATGGAGACCCCTTGGGGAGCAGGCTCGGG	6
T0/7	GAACCCTCGGTGCCCTAACATTGACCTCGTCTACCTCTGGGGAACCCCTCGTCCGAGCCC	7870
	LGATGL*	
0	CTCCCCAGCCCCATCCCAAACCCTTCAGTGCCAAAAACAACAACAAAATAGAAAGCGCAA	0
7707	GAGGGGTCGGGGTAGGGTTTGGGAAGTCACGGTTTTTTGTTGTTGTTTTTATCTTTCGCGTT	7680
000		<b>6</b>
1007	GGTGGTGGTTGGTGGTGTTCTTCCTACTAAGTTGTCCAAAAGGACTTCTTAACT	7340
1,00	AAAACCATTTTGCTGTCCCTTTTTCCTTTTGATGTTCTTTCACCCTTTTCTGTTTGCTA	
T # C 7	TTTTGGTAAAACGACAGGGAAAAAAAAACTACAAGAAAGTGGGAAAAGACAAACGAT	2000
1006	AGTGAGGATGAAAAATAACACTGTACTGCAATAAGGGGAGAGTAACCCTGTCTAATGAA	
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6	TTTTTCTTACTAATATCCATGG		1015
31.20	CTCCTTTGACGTGACAAATAA	TGGACACAGAGACTCTCATCTCAGTGACCTTGTGATTACTCCTTTGACGTGACAAAATAA	1
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ч	GAATTCCGTGAGTGCATGGGAGGGTGCTGAATATTCCGAGACACTGGGACCACAGCGGCA 	9
61	GCTCCGCTGAAAACTGCATTCAGCCAGTCCTCCGGACTTCTGGAGCGGGGACAGGGCGCAAAAAAAA	120
121	GGGCATCAGCAGCCAGCAGGACCTGGGAAATAGGGATTCTTCTGCCTCCACTTCAGG +++	180
181	TTTTAGCAGCTTGGTGCTAAATTGCTGTCTCAAAATGCAGAGGATCTAATTTGCAGAGGA +++++	240
241	AAACAGCCAAAGAAGGAAGGAGAAAAAGGAAAAAAAAAA	300
301	TACTTTTCTTGGAAATGCAAAAGATTATGCATATTTCTGTCCTCCTTTCTCCTGTTTTAT ++++	360
	M Q K I M H I S V L L S P V L W	1
361	GGGGACTGATTTTTGGTGTCTCTTCTAACAGCATACAGATAGGGGGGGCTATTCCTAGGG ++++++	420

## F16. 2B

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660 GACATTTATGGTAGTGTAAAAACGCCTTGTGAGGTGCAGAGGAAGTAGTGAGGGTCGA CTGTAAATACCATCACATCATTTTGCGGAACACTCCACGTCTCCTTCATCACTCCAGCT 601

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ı	FYILVGGLGAMLVALIEFC-
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3001 -----+---+----+ 3060 GTTCCGACAGTTAATGTCCTTCATGACCTCTTTTACCTGCACAATACTGAGGTCTTAAAG CAAGGCTGTCAATTACAGGAAGTACTGGAGAAAATGGACGTGTTATGACTCCAGAATTTC

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	TLDSGSTKEFFRRSKIAVYE	1
2161	aaaatgtggtcttacatgaaatcagcggagccatctgtgtttaccaaaacaacagcagac	
T 0 T 7	ttttacaccagaatgtactttagtcgcctcggtagacacaaatggttttgttgtcgtctg	2220
	KMWSYMKSAEPSVFTKTTAD	1
2221	ggagtggcccgagtgcgaaagtccaagggaaagttcgccttcctgctggagtcaaccatg	0
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#### F16. 31

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2640 2700 2760 aagaacacccaaaactttaagcctgctcctgccaccaacactcagaattatgctacatac gtggctttgatagaattctgttacaaatcacgggcagagtccaaacgcatgaaactcaca caccgaaactatcttaagacaatgtttagtgcccgtctcaggtttgcgtactttgagtgt ttcttgtgggttttgaaattcggacgaggacggtggttgtgagtcttaatacgatgtatg tctcttccgatgttgcacataccttgtctctcacaattctagatccctagggaaccttaa agagaaggctacaacgtgtatggaacagagagtgttaagatctagggatcccttggaatt X · X K z æ ø × S Н Z EI A × H > ĸ Ø S Ŋ Д E × G ပ ы ч ď, 臼 × ø 2581 2641 2701

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TAGARACCTAAGATACTGGTC  I F G F Y D Q  CACACATCCTTTGTTACGCCT  421++++++	_		S N S F S V T N A F C S Q F S R G V Y A - ATCTTTGGATTCTATGACACACACCCTGACCTCCTTCTGTGGGGCCCTG  361+++ 420

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### F16. 4F

	GAGAATCGGACCATAGTAGTGACTACCATTCTGGAATCACCATATGTAATGTACAAGAAG	CGGACCATAGTAGTGACTACCATTCTGGAATCACCATATGTAATGTACAAGAAG		1380
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	TIGGIACTCGTTGACCTTCCTTTACTTGCTATACTTCCGATAACACATCTGGATCGGATA	TGCTATACTTCCGATAACA		1440
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	CCAC	ATACCTTGCCGTACCAACCC		1.560
ત	GARDPETKI	W N G W V	ELVYG	i
	AGAGCTGATATAGCTGTTGCTCCACTCACTATAACATTGGTCCGTGAAGAAGTCATAGAT	CACTATAACATTGGTCCGT		(
	TCTCGACTATATCGACAACGAGGTGAGTGATATTGTAACCAGGCACTTCTTCAGTATCTA	AGTGATATTGTAACCAGGCA		1.620
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ъ		C D I S P R S L S G R I V G G V W W F F	;
	1981	ACCCTGATCATAATTTCTTCCTATACTGCCAATCTCGCTGCTTTCCTGACTGTGGAGAGG	0
	† ) ;	TGGGACTAGTATTAAAGAAGGATATGACGGTTAGAGCGACGAAAGGACTGACACCTCTCC	2040
ಗ		TLIIISSYTANLAAFLTVER.	i
	1,400	<b>ATGGTTTCTCCCATAGAGTGCTGAAGACTTAGCTAAACAGACTGAAATTGCATATGGG</b>	
	7 7 7 T	TACCAAAGAGGGTATCTCTCACGACTTCTGAATCGATTTGTCTGACTTTAACGTATACCC	2100
ರ		M V S P I E S A E D L A K Q T E I A Y G	·
	1016	ACCCTGGACTCCGGTTCAACAAAGAATTTTTCAGAAGATCCAAAATTGCTGTGTACGAG	,
	7 7 7 7	TGGGACCTGAGGCCAAGTTGTTTTCTTAAAAGTCTTCTAGGTTTTTAACGACACATGCTC	2160
ď		TLDSGSTKEFFRRSKIAVYE -	í
	2161	AAAATGTGGTCTTACATGAAATCAGCGGAGCCATCTGTGTTTACCAAAACAACAGCAGAC	
	1017	TTTTACACCAGAATGTACTTTAGTCGCTCGGTAGAAATGTTACAAAACGTTAGAAAACAAAA	2220

#### F/G 41

í	DKGECGAKDSGSKDKTSALS	ਰ
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2520	GATAAGGGGGAATGTGGAGCCAAGGACTCCGGGAGTAAGGACAAGACCAGCGCTCTGAGC	
i	AVLKLSEQGILDKLKNKWWY	a
00 #		
24.60	GCAGTATTGAAACTCAGTGAACAAGGCATCTTAGACAAGCTGAAAAAAAA	
•	SKGYGVATPKGSALGTPVNL	್ರಡ
.400 .400	AGGTTTCCGATACCACACCGTTGGGGATTTCCGAGTCGTAATCCTTGCGGACATTGGAA	
6	TCCAAAGGCTATGGTGTGCCAACCCCTAAAGGCTCAGCATTAGGAACGCCTGTAAACCTT	
;	NEYIEQRKPCDTMKVGGNLD	æ
0.4 0.4 0.4		
6	AATGAGTACATTGAGCAGAAAACCATGTGATACGATGAAAGTTGGTGGAAATCTGGAT	
i	GVARKSKGKFAFLLESTM	æ
7.7	CCTCACCGGGCTCACGCTTCAGGTTCCCTTTCAAGCGGAAGGACGACCTCA	
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# F1G. 4J

2580	1		7640	ı	2700	ı	2760		ı		7820
CTGAGCAATGTGGCAGGCGTTTTCTATATACTTGTCGGAGGTCTGGGGCTGGCCATGATG 2521+++++++-	LSNVAGVFYILVGGLGLAMM	GTGGCTT	++++++++	VALIEFCYKSRAESKRMKLT	AAGAACACCCAAAACTTTAAGCCTGCTCCTGCCACCAACACTCAGAATTATGCTACATAC  2641+++++++	K N T Q N F K P A P A T N T Q N Y A T Y	AGAGAAGGCTACAACGTGTATGGAACAGAGTGTTAAGATCTAGGGATCCCTTCCCACT	TCTCT	REGYNVYGTESVKI*	GGAGGCATGTGAGAAAATCACCGAAAACGTGGCTGCTTCAAGGATCCTGAGCCAG	CCTCCGTACACTACTCTCTTTAGTGGCTTTTGCACCGACGAGGTTCCTAGGACTCGGTC
	ત્ત			ď		ત			๙		

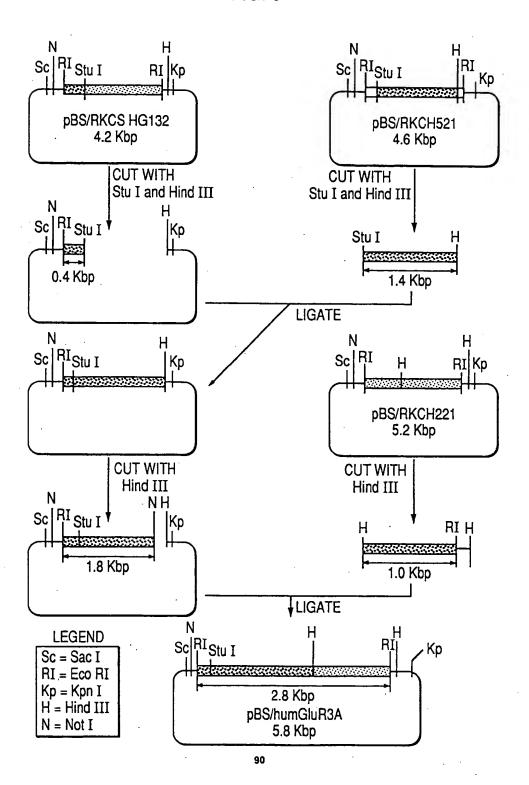
## F16 4K

	TTAGGAATTC 1+ 3070	3061
2000		
0,90%	AACCAACAAAAATGGACATGCATCAAACCCTTGATGTATTAATATTTATATATTTCA	3001
2000	TGTTACGTTTGGTAGTAACTTTAGAAAAACGAAACGAACTTTTTTTT	1 ,
0	•	141
0467	TTATCCITTTGACTAAAAAAAAGGAAGTCACGGAATACCTTGTGAGACTCTGAGCGC	)
		7881
7880	TAAAGTGAGGAACCACACCCGTACTGTGCTTATAACGACTACCACGTTACTGGAAAG	707
0		1696

#### F/6 5

.. GSALGNAVNLAVLKLNEQGLLDKLKNKWWYDKGECGSGGDSKDKT.. GluR3A GluR3B

FIG. 6



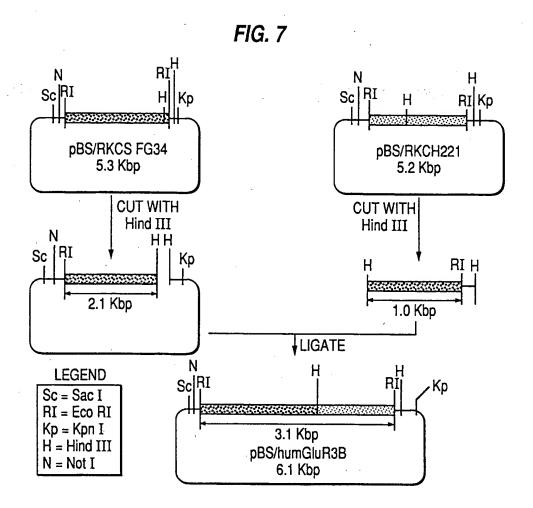


FIG. 8

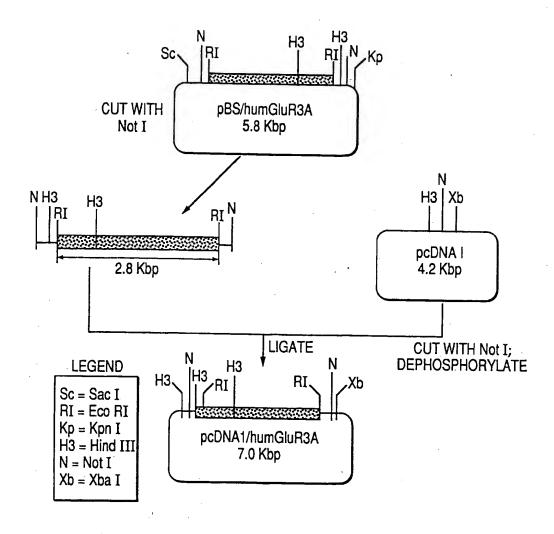
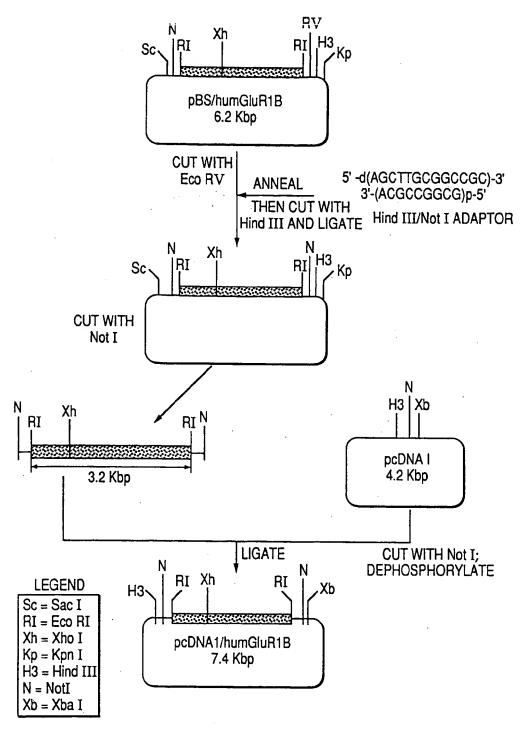


FIG. 9



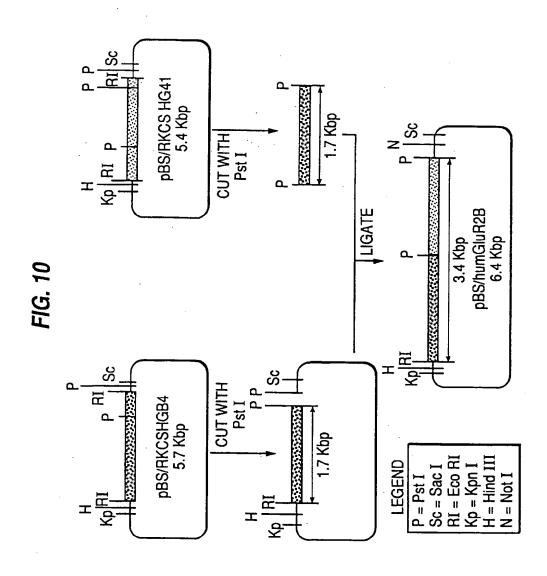
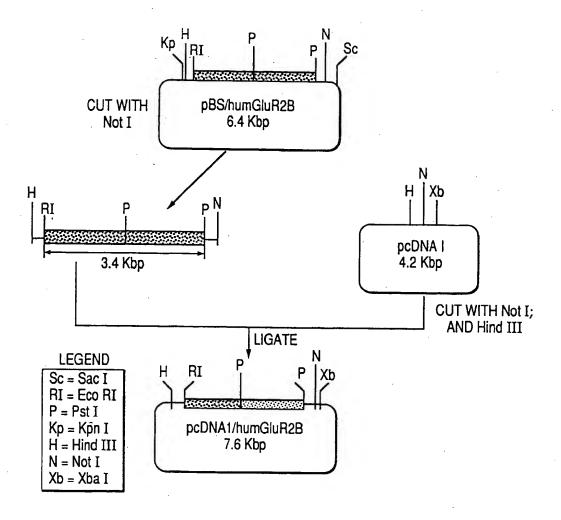
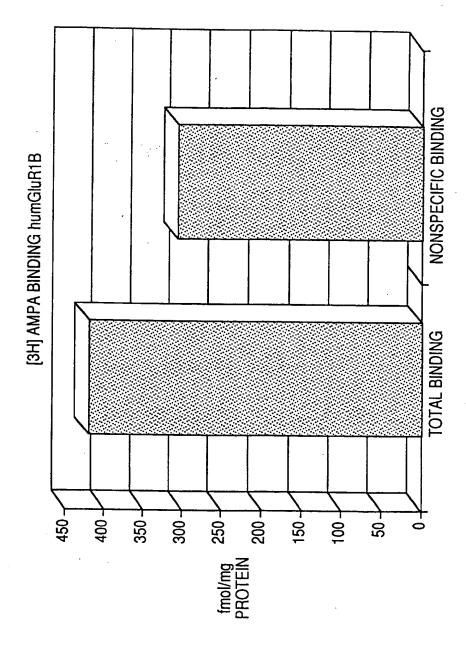
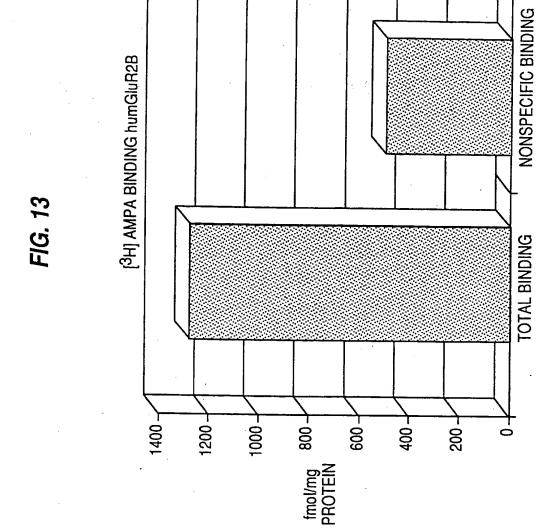


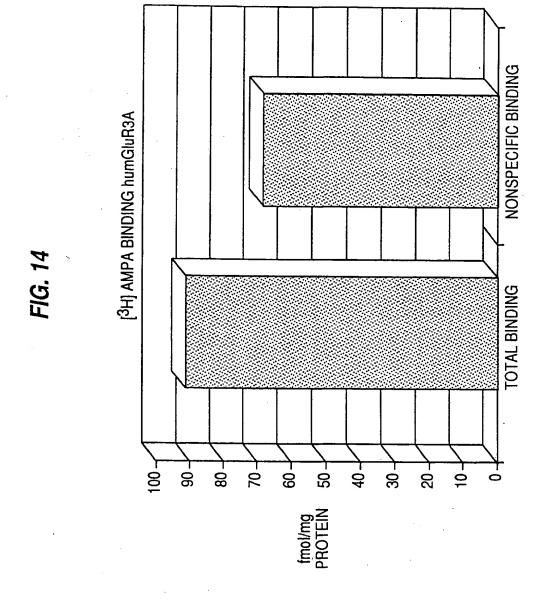
FIG. 11

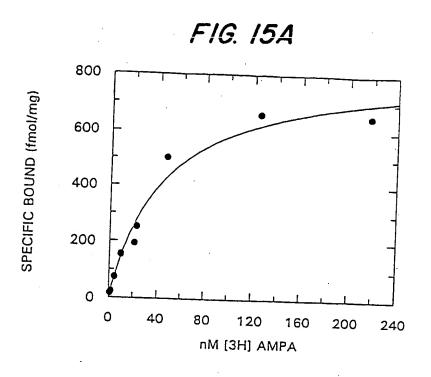


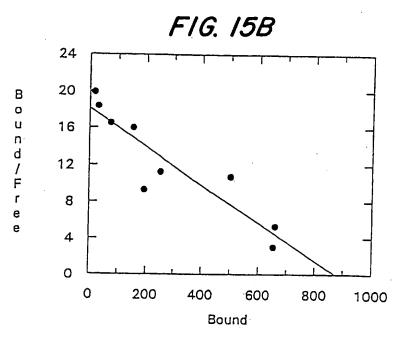


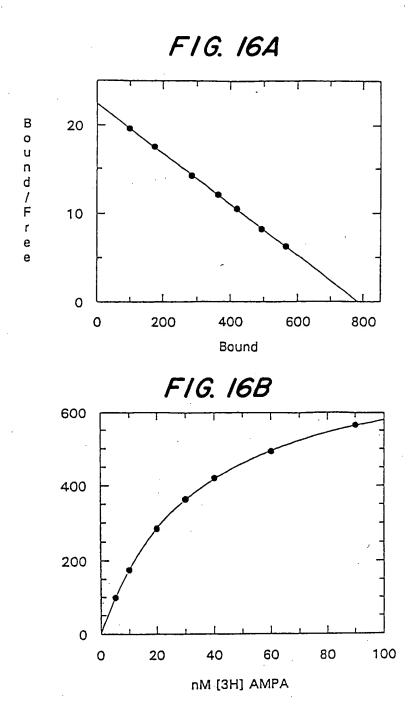


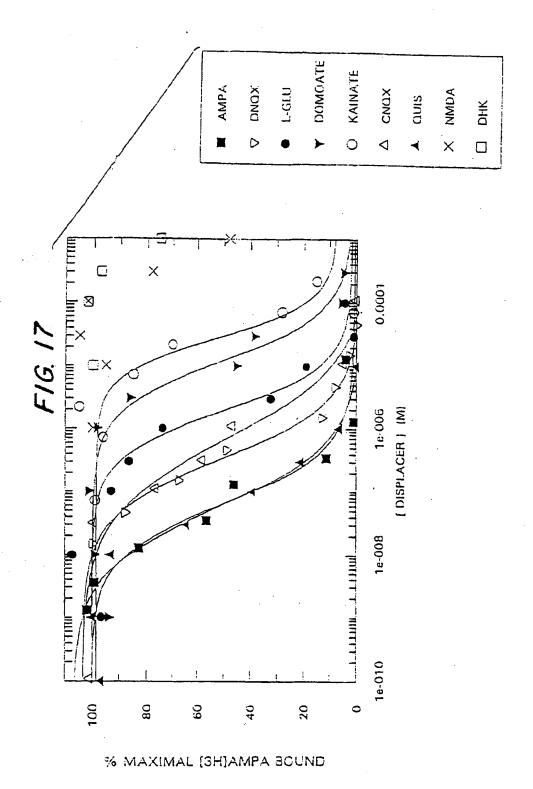












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(12)

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(54) Amino-hydroxy-methyl-isoxazole-propionate binding human glutamate receptors.

(5) Described herein are isolated polynucleotides which code for a family of AMPA-type human CNS receptors. The receptors are characterized structurally and the construction and use of cell lines expressing these receptors are disclosed.



EP 0 574 257 A3



#### **EUROPEAN SEARCH REPORT**

Application Number EP 93 30 4500

Category	Citation of document with in of relevant pas		Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.CLS)		
Y	WO-A-91 06648 (THE S BIOLOGICAL STUDIES) * the whole document		1-18	C07K15/00 C12N15/12 C12N5/08 C12P21/02		
Y,D	SCIENCE., vol.249, no.4968, 19 pages 556 - 560 KEINÄNEN K. ET AL '/ * the whole document		1-18	C12P21/08 G01N33/48		
<b>A</b>		EQUENCING AND MAPPING,	1-12			
				TECHNICAL FIELDS SEARCHED (Int.CL.5) CO7K		
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	The present search report has be	nen drawn un for all deine				
	Place of sourch	Date of completion of the search	<u> </u>	Prostor		
BERLIN  CATEGORY OF CITED DOCUMENTS  X: particularly relevant if taken alone Y: particularly relevant if combined with another document of the same extegory A: technological background		T: theory or princip E: earlier patent do after the filling d	T: theory or principle underlying the inves E: earlier patent document, but published after the filling date D: document cited in the application L: document cited for other reasons			